



# Down-regulation of microglial activation may represent a practical strategy for combating neurodegenerative disorders

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**Summary** Chronic neurodegenerative disorders are characterized by activation of microglia in the affected neural pathways. Peroxynitrite, prostanoids, and cytokines generated by these microglia can potentiate the excitotoxicity that contributes to neuronal death and dysfunction in these disorders – both by direct effects on neurons, and by impairing the capacity of astrocytes to sequester and metabolize glutamate. This suggests a vicious cycle in which the death of neurons leads to microglial activation, which in turn potentiates neuronal damage. If this model is correct, measures which down-regulate microglial activation may have a favorable effect on the induction and progression of neurodegenerative disease, independent of the particular trigger or target involved in a given disorder. Consistent with this possibility, the antibiotic minocycline, which inhibits microglial activation, shows broad utility in rodent models of neurodegeneration. Other agents which may have potential in this regard include PPAR $\gamma$  agonists, genistein, vitamin D, COX-2 inhibitors, statins (and possibly policosanol), caffeine, cannabinoids, and sesamin; some of these agents could also be expected to be directly protective to neurons threatened with excitotoxicity. To achieve optimal clinical outcomes, regimens which down-regulate microglial activation could be used in conjunction with complementary measures which address other aspects of excitotoxicity.

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## Chronic microglial activation as a mediator of neurodegeneration

There is growing evidence that activated microglia play a key pathogenic role in chronic neurodegenerative disorders as well as in the tissue damage consequent to stroke or brain trauma [1–8]. For reasons that remain largely obscure, the death or dysfunction of neurons typically results in activa-

tion of neighboring microglia. When activated, these microglia become a prominent source of oxidants, prostanoids, and inflammatory cytokines; this in turn can promote death and dysfunction of neurons, resulting in a vicious cycle in which a progressive loss of neurons is accompanied and abetted by sustained microglial activation.

The chief mediator of the pathogenic impact of activated microglia appears to be peroxynitrite [9]. This arises owing to activation of microglial NADPH oxidase – a potent generator of superoxide that in the brain is expressed primarily in microglia

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– in conjunction with induction of the inducible isoform of nitric oxide synthase (iNOS); the superoxide and nitric oxide (NO) produced by these enzymes can react avidly and spontaneously to yield the potent oxidant peroxynitrite, which readily diffuses through cell membranes, and thus can attack neighboring neurons and astrocytes. Superoxide per se does not readily penetrate cell membranes, and its product hydrogen peroxide is a relatively weak oxidant (except in the presence of free iron or copper). NO per se is only toxic at high concentrations. Thus, superoxide and NO are much more toxic jointly than separately, as they give rise to peroxynitrite. A central role for peroxynitrite in the pathogenicity of activated microglia is consistent with evidence that inhibition or diminished expression of either NADPH oxidase or iNOS substantially reduces the neurotoxicity of activated microglia in vitro, and diminishes the severity of rodent neurodegenerative syndromes in vivo [10–20].

In neurodegenerative conditions, neuronal death, whether from necrosis or apoptosis, is typically associated with, and mediated by, excessive free intracellular calcium (induced by excitotoxic glutamate exposure), oxidant stress, and mitochondrial dysfunction (leading to ATP deficit and/or superoxide production) [21–25]. Glutamate-mediated excitotoxicity appears to play some role in most acute or chronic neurodegenerative conditions, and oxidant stress, as well as inefficient bioenergetics (owing to mitochondrial failure or ischemia, for example), can markedly potentiate the toxicity of glutamate to neurons [10,23,26]. In particular, peroxynitrite derived from microglia has been shown to boost cell death in glutamate-exposed neurons in vitro [10]. The basis of this effect is still unclear, although peroxynitrite attack on the mitochondrial respiratory chain – most notably complexes I and II [27–30] – as well as oxidant damage to membrane ion transporters [31,32], likely contribute. ATP deficit impairs the mechanisms which expel and sequester free intracellular calcium – thus exacerbating calcium overload – and can promote necrotic or apoptotic cell death by additional mechanisms. In vivo, microglial-derived peroxynitrite also promotes excitotoxicity by inhibiting the transport mechanisms by which astrocytes sequester extracellular glutamate [20,33,34]. Thus, peroxynitrite promotes excitotoxicity both by increasing the exposure of neurons to glutamate, and by increasing the sensitivity of neurons to this neurotransmitter.

It should be noted that activated microglia are not the only source of peroxynitrite in neurodegenerative conditions. The excess in intracellular free calcium associated with excitotoxicity strongly

activates the neuronal isoform of nitric oxide synthase (nNOS), expressed in a high proportion of neurons [21,35]. Furthermore, nNOS is often partially “uncoupled”, owing to suboptimal intracellular levels of arginine and/or tetrahydrobiopterin; this means that the calcium-activated enzyme produces superoxide as well as NO, enabling production of peroxynitrite within neurons [36–38]. Dysfunctional neural mitochondria can also act as a source of superoxide. This presumably explains why nNOS knockout mice tend to be less sensitive to ischemia or neurotoxins [35,39,40]. The implication is that suppression of microglial activation may only partially alleviate the pathogenic impact of peroxynitrite in neurodegenerative disorders.

The inducible form of cyclooxygenase (COX-2) is also induced in activated microglia [41,42]. COX-2-derived prostanoids are somewhat analogous to peroxynitrite in regard to their impact on excitotoxicity: they act directly on neurons to increase their susceptibility to glutamate-induced death, and they induce astrocytes to extrude rather than sequester glutamate [43–47]. Although COX-2 is minimally expressed in healthy neurons, it can be induced in neurons undergoing excitotoxicity [45]; the resulting increase in prostanoid production can have a feed-forward impact on glutamate toxicity.

Activated microglia also produce a range of inflammatory cytokines, including IL-1 and TNF- $\alpha$ . The latter has been reported to potentiate glutamate-mediated neurotoxicity – although a contrary finding has also been reported [48,49]. These cytokines can also suppress glutamate uptake and metabolism by astrocytes [50–53]. Thus, microglial-derived cytokines may contribute to excitotoxic neurodegeneration – although the evidence for this is less clear-cut than in the case of peroxynitrite or prostanoids.

The proportion of brain microglia which are activated tends to increase as a function of aging; this is observed even in animals and humans that have not been traumatized and who do not suffer from known neurodegenerative disease [54–56]. This phenomenon might help to explain why chronic neurodegenerative disorders are far more common in the elderly. Furthermore, microglial activation may become self-sustaining; activated microglia are observed in the substantia nigra of monkeys fully a year after a single administration of MPTP [57]. A similar phenomenon has been noted when humans transiently exposed to MPTP are autopsied [58]. It is not clear whether this prolonged activation might stem from autocrine mechanisms, or whether it is sustained by the continuing death

and dysfunction of neurons damaged by the activated microglia.

### Activated microglia in Parkinson's and Alzheimer's diseases

In Parkinson's disease, microglial activation is prominent in the substantia nigra [21,4,59]. Dopaminergic neurons, high in iron and low in glutathione, appear to be unusually sensitive to oxidant stress [60]; moreover, the healthy substantia nigra hosts a relatively high concentration of microglia. It thus is not surprising that a continuous intracerebral infusion of lipopolysaccharide (LPS) in rats, resulting in activation of microglia, leads to selective loss of dopaminergic neurons in the substantia nigra [61,62]; a similar effect has been reported following thrombin infusion [63]. This effect can be replicated in vitro – when dopaminergic neurons are co-cultured with microglia, addition of LPS leads to death of the neurons [18]. Microglial oxidant production appears to be a key mediator of this effect, since inhibitors of NADPH oxidase, or the use of microglia in which this enzyme complex is genetically defective, prevents neuron death. Analogously, the ability of rotenone or of MPTP to promote degeneration of dopaminergic neurons in vivo or in vitro is substantially suppressed when NADPH oxidase activity is concurrently inhibited or is genetically defective [14,15,17]. The inducible nitric oxide synthase (iNOS), prominently expressed in activated microglia, also appears to contribute to oxidant-mediated death of dopaminergic neurons, since pre-administration of iNOS inhibitors, or use of mice genetically deficient in this enzyme, protects rodents from MPTP- or LPS-induced parkinsonism [11,64,62]. These findings are clearly consistent with the possibility that microglia-derived peroxynitrite is a key pathogenic factor in Parkinson's disease.

With respect to Alzheimer's disease, microglial activation is likewise prominent in affected brain regions in this disorder [3,65], and it has been observed that  $\beta$  amyloid-42 is far more toxic to neurons in vitro when they are co-cultivated with microglia [13]. This toxicity is markedly blunted when the microglia used in this study are derived from NADPH oxidase-deficient mice – consistent with the fact that  $\beta$  amyloid strongly activates this enzyme in microglia [66]. Moreover, cytokines produced by activated microglia can both stimulate neuronal production of  $\beta$  amyloid precursor protein, and promote its conversion to  $\beta$  amyloid [67–69]; COX-2 products likewise up-regulate pro-

duction of  $\beta$  amyloid [69,70]. This suggests a vicious cycle in which neural production of  $\beta$  amyloid leads to microglial activation, which in turn promotes neuronal cell death while further stimulating  $\beta$  amyloid production [3].

### Broad efficacy of minocycline confirms a pathogenic role for microglia

The antibiotic minocycline, which readily penetrates the blood-brain barrier, has been shown to suppress microglial activation triggered by a broad range of activating stimuli; furthermore, it can do so in concentrations that are close to the clinical range for this well-tolerated drug [71–76]. The biochemical basis of this effect is not clear, although prevention of p38 MAP kinase activation appears to play a key role in this regard [71,74]; p38 signaling plays a central role in microglial activation [77]. Thus, it is of particular interest that pre-administration of minocycline has been shown to protect rodents from a wide range of neurodegenerative conditions, including rodent models of Parkinson's and Huntington's diseases, ALS, multiple sclerosis, stroke, excitotoxicity, and brain trauma [75,76]. These findings may be interpreted as strongly suggestive evidence that microglial activation is a prominent mediator of the neural death and dysfunction that characterizes these syndromes. Moreover, they have encouraged clinical efforts to evaluate minocycline as a neuroprotective agent in a range of disorders; for example, the impact of minocycline therapy on the progression of Parkinson's disease is currently being studied.

In addition to minocycline, a variety of other agents and strategies have the potential to down-regulate microglial activation, and thus possibly provide protection from a range of neurodegenerative disorders. Some of these are drug-related strategies – such as minocycline – that could most appropriately be applied in patients who are in the early stages of neurodegenerative disorders, or who are at very high genetic risk for same. Other strategies, involving nutrients and food factors, might reasonably be included in lifestyle regimens for healthy people who wish to preserve effective brain function to a ripe old age.

### PPAR $\gamma$ agonists

Microglia express the PPAR $\gamma$  transcription factor, and agonists for this receptor, such as pioglitazone,

inhibit LPS-triggered induction of iNOS and of TNF- $\alpha$  in microglial cell culture [78–80]. Increased expression of I $\kappa$ B- $\alpha$ , which inhibits activation of the NF- $\kappa$ B transcription factor, may mediate this effect [81] – although contrary evidence has also appeared [78]. In vivo, pioglitazone pre-treatment protects dopaminergic neurons in the substantia nigra of mice treated with MPTP; it is somewhat less effective in preventing loss of dopaminergic terminals in the striatum [81,82]. Pioglitazone and other PPAR $\gamma$  agonists also have a favorable impact on the processing of  $\beta$  amyloid precursor protein, reducing the expression of the  $\beta$ -secretase required for production of  $\beta$  amyloid; they also diminish Alzheimer's-like pathology in transgenic mice which overexpress the  $\beta$  amyloid precursor protein [69,83,84]. Moreover, this drug is effective in experimental autoimmune encephalomyelitis, a rodent model of multiple sclerosis, and an anecdotal report of apparent response to this agent in an MS patient has appeared [85,86]. Given the fact that pioglitazone is a well tolerated drug, this agent merits further clinical evaluation in neurodegenerative syndromes.

PPAR $\gamma$  agonists may also have direct effects on certain neurons that protect against excitotoxicity. In cultured cerebellar granule neurons, administration of troglitazone up to 2 h following glutamate exposure was protective, even though the elevation of intracellular free calcium was not influenced; evidently, PPAR $\gamma$  influences a downstream event triggered by calcium overload [87].

## Genistein

Parkinson's disease and ALS appear to be more common in men than in women; furthermore, epidemiological studies suggest that early menopause may increase risk for Parkinson's disease, whereas postmenopausal estrogen replacement may reduce this risk [88–91]. Thus, it is notable that estrogen exerts anti-inflammatory effects on microglia, acting via either isoform of the estrogen receptor [92,93]; this may rationalize the utility of estrogen therapy in rodent models of Parkinson's diseases [94–97]. With respect to Alzheimer's disease, epidemiology has pointed to a protective role for postmenopausal hormone replacement, whereas estrogen administration to elderly women in prospective studies has not shown such protection; the discrepancy between these results remains to be explained [98,99].

Although both isoforms of the estrogen receptor are expressed in the brain, the expression of

ER $\beta$  is broader and more prominent; the brains of ER $\beta$  knock-out mice show marked abnormalities [100]. In light of the fact that hippocampal neurons are targeted in dementia, it is notable that ER $\beta$  is the predominant isoform in the primate hippocampus [101,102]. Microglia express ER $\beta$ , and selective agonists for this receptor exert anti-inflammatory effects on microglia, suppressing LPS-mediated induction of both iNOS and COX-2 [93]. Genistein, a potent and selective agonist for this receptor in physiologically achievable concentrations, is protective in rodent models of ALS and stroke [103]. Furthermore, in the low nanomolar free concentrations that can be achieved clinically, genistein alleviates the cytotoxicity of  $\beta$  amyloid to a neuroblastoma-derived cell line as well as cultured hippocampal neurons; this effect presumably reflected interaction with neuronal estrogen receptors, however [104,105]. Genistein likewise can alleviate the dopaminergic neurodegeneration evoked by LPS in rat mesencephalic-glia cultures; however, high nanomolar concentrations were required for this effect, suggesting that tyrosine kinase inhibition may have been responsible [106]. Further evaluation of genistein in rodent models of neurodegeneration appears warranted. Physiologically significant intakes of genistein have the potential to provide a range of protective health benefits, owing to the ability of this agent to activate ER $\beta$  at concentrations that have minimal impact on the cancer-promoting ER $\alpha$  receptor [107]. An oral daily intake of 54 mg genistein has demonstrated beneficial clinical effects in recent studies [108,109]; this would be supplied by about 150 mg of mixed isoflavone glycosides extracted from soy.

ER $\beta$  can also provide direct protection from excitotoxicity in some neural pathways. Thus, specific agonists for ER $\beta$  have been shown to induce the anti-apoptotic protein Bcl-2 in hippocampal neurons; these agents protect hippocampal neurons from glutamate-mediated excitotoxicity in vitro, and from brief global ischemia in vivo [110–113]. The protective impact of estrogen on MPTP-treated rats appears to be mediated by ER $\alpha$  receptors [97]; however, another study suggests that ER $\beta$  is responsible for the favorable influence of estrogens on cultured dopaminergic neurons exposed to MPP(+) [114].

## Vitamin D

Microglial cells express the vitamin D receptor, and calcitriol inhibits expression of iNOS by microglial cells exposed to LPS and other activating agonists

[115–117]. This may reflect the presence of a vitamin D response element in the promoter of the iNOS gene. Furthermore, calcitriol boosts astrocyte production of glial-derived neurotrophic factor (GDNF), a growth factor that provides particular protection for dopaminergic neurons of the substantia nigra [118,119]. In rats, calcitriol administration has a protective effect in 6-hydroxydopamine-induced Parkinsonism as well as in experimental autoimmune encephalomyelitis [120,121].

1 $\alpha$ -Hydroxylase, the enzyme which converts 25-hydroxyvitamin D to the active hormone calcitriol, is expressed by activated but not quiescent microglia [122]. Thus, activated microglia generate calcitriol when incubated with 25-hydroxyvitamin D. This raises the intriguing possibility that autocrine production of calcitriol by microglia could be boosted by improving vitamin D status; supplemental or autogenous vitamin D might have the potential to increase microglial and astrocyte exposure to calcitriol during the early stages of neurodegenerative syndromes – without entailing the hypercalcemic risk evoked by direct calcitriol administration. In this regard, incidences of both Parkinson's disease and ALS have been found to correlate positively with latitude [123–125]; no such relationship has been reported for Alzheimer's disease, however. High but tolerable doses of vitamin D – rather than calcitriol – should be studied in various rodent models of neurodegenerative disease. In humans, daily doses as high as 10,000 IU could reasonably be tested, inasmuch as physiological capacity for *uv*-catalyzed endogenous production of vitamin D is on the order of 10–20,000 IU daily [126].

## COX-2 inhibitors

Individuals who have used NSAIDs chronically for years appear to be at substantially lower risk for both Parkinson's disease and Alzheimer's; this pertains to aspirin as well, but only when used in high anti-inflammatory doses [127,128]. This suggests that prostanoids derived primarily from COX-2 in activated microglia may act as mediators of neurodegeneration; indeed, as noted above, COX-2 products can sensitize neurons to excitotoxicity, while also impairing the ability of astrocytes to sequester glutamate [43–47]. The possibility that these prostanoids also act, directly or indirectly, to sustain microglial activation, is suggested by the observation that activated microglia are less common in the brains of humans or rodents that have been treated chronically with NSAIDs [55]. In vitro,

COX inhibitors suppress expression of iNOS in LPS-activated microglia; perplexingly, PGE<sub>2</sub> boosts this expression [129]. Supernatants derived from an microglial-like cell line (THP-1) activated with LPS are cytotoxic to neuroblastoma-derived cells in vitro; this cytotoxicity is largely alleviated if the THP-1 cells are incubated with COX inhibitors [130]. COX-2 inhibitors are protective in the MTPT model of Parkinsonism in mice, as well as in rodent models of Alzheimer's and ALS [131–133]. Although the increased cardiovascular risk associated with COX-2 inhibitor therapy has recently discouraged the use of these drugs [134], it seems likely that they would be no more risky than non-specific COX inhibitors if used in conjunction with low-dose aspirin to stabilize platelets. (Indeed, they might be safer, since the COX-1 activity of vascular endothelium would be largely preserved.) Thus, the use of COX-2 inhibitors + low-dose aspirin should be considered as a clinical neuroprotective strategy.

Diets high in fish and fish oil have been associated epidemiologically with decreased risk for Alzheimer's disease [135–137]. Could this reflect modulation by  $\omega$ -3 fats of COX-2-mediated prostanoid production? Whether fish intake might influence risk for Parkinson's disease appears to have received little attention.

## Statins – and policosanol?

Several – though not all [138,139] – case-control studies have concluded that patients who use statins may be at decreased risk for Alzheimer's disease [140–143]. These findings are subject to the bias that people who seek out and use medical care may tend to be more mentally competent than those who do not; however, one study noted that use of other types of lipid-lowering agents was not associated with protection in this regard [141]. Nor is high cholesterol per se protective – quite to the contrary, elevated LDL cholesterol may be a risk factor for dementia [144–147]. The impact of statin use on Parkinson's risk has apparently received little attention, although one study failed to note any evident impact of on-going statin therapy on the clinical course of the disease [148].

Evidence that statin therapy might reduce Alzheimer's risk has motivated several groups of researchers to examine the impact of statins on microglial activation. In vitro, statins suppress the rac1-dependent activation of NADPH oxidase and induction of iNOS in  $\beta$  amyloid-stimulated microglia [149]; LPS-mediated induction of iNOS

and of cytokines is also inhibited, as is secretion of apoE (which promotes  $\beta$  amyloid fibrillogenesis and deposition) [150,151]. These effects appear to be mediated by decreased isoprenylation of microglial signaling proteins, as they can be reversed by addition of geranylgeranyl pyrophosphate. There is, however, one discordant report, observing that statins themselves can have an activating impact on microglia [152]. There is also some evidence that statins may also have a favorable influence on the processing of amyloid precursor protein, such that  $\beta$  amyloid secretion is suppressed [153–155]; this effect may be mediated, in part, by reduced cellular cholesterol levels. Moreover, there are two reports that atorvastatin diminishes glutamate-mediated excitotoxicity in cortical neuron cultures [156,157]. Whether such effects can be achieved in vivo with tolerable clinical doses of statins, is not yet clear. Statins may vary with respect to their access to the brain; atorvastatin does not cross the blood-brain barrier [158]. Statins can also influence brain function by up-regulating endothelial expression of eNOS – an effect that could help to maintain efficient brain perfusion and thereby aid brain bioenergetics [159–161]. Statin pre-treatment decreases infarct volume following focal ischemia in rodents [162].

If statins do indeed have neuroprotective potential, it will be important to determine whether policosanol – a mixture of non-toxic sugar cane waxes which lowers LDL cholesterol by down-regulating expression of HMG-CoA reductase – likewise can be protective in this regard [163–165]. Whether policosanol can influence expression of this enzyme in the brain has not been determined. It is encouraging to note, however, that policosanol appears to share the osteoprotective properties of statins [166]; thus, its physiological effects may be parallel to those of statins. Moreover, hexacosanol acid, a component of policosanol, is reported to protect rats from central kainic acid-mediated excitotoxicity after intraperitoneal administration [167]. The particular merit of policosanol is that, whereas excessive concentrations of statins can induce severe toxicity by over-inhibiting HMG-CoA reductase in skeletal muscle, even very high concentrations of policosanol do not appear to reduce expression of this enzyme by more than about 50% [163]; this likely accounts for the non-toxicity of this agent in animal studies, and its excellent tolerability in clinical trials [168]. Thus, if policosanol proves to have neuroprotective activity, it would be feasible for the healthy general population to use this agent – without the regular physician monitoring that statin use entails.

## Caffeine

Regular coffee drinkers are at markedly lower risk for Parkinson's disease [169], and two epidemiological studies suggest that Alzheimer's disease may also be less common in coffee drinkers [170,171]. Caffeine has well documented neuroprotective effects in a range of rodent models, including those for Parkinson's disease, stroke, and excitotoxicity [172]; moreover, caffeine is reported to decrease the toxicity of  $\beta$  amyloid to cultured cerebellar neurons in vitro [173]. This protection appears to reflect inhibition of adenosine type 2A receptors (A2A), widely expressed in the brain [172]. In particular, such receptors are found on microglial cells, and selective agonists for this receptor promote induction of COX-2 in these cells [174]. Thus, it is conceivable that caffeine neuroprotection reflects, at least in part, down-regulation of COX-2 expression in microglia. However, the main impact of A2A antagonists on neurodegeneration may reflect a down-regulation of glutamate release from excitatory synapses; evidently, adenosine plays a physiological role in promoting the extracellular glutamate excess that mediates excitotoxicity [175–178].

Importantly, whereas cardiovascular responses to caffeine tend to rapidly down-regulate, the protection afforded by caffeine in MPTP-induced Parkinsonism was seen in rats that had been chronically pre-treated with caffeine [179]. In light of the fact that the impact of caffeine on cardiovascular risk is equivocal, and heavy coffee use has been linked to reduced risk for diabetes, those who enjoy their daily coffee can take comfort in the thought that they may be protecting their brain in the process.

## Cannabinoids

Cannabinoids, acting via CB1 or CB2 receptors expressed by microglial cells, inhibit LPS-mediated induction of iNOS in microglia [180,181]; they also inhibit activation of microglia by  $\beta$  amyloid, in vitro and in vivo, and prevent the cognitive dysfunction and neuronal death induced by intracerebral  $\beta$  amyloid administration in rats [182]. In addition, cannabinoids directly protect neurons from glutamate-mediated excitotoxicity, in vitro and in vivo [181,183–188]; moreover, like A2A antagonists, they act on excitatory pre-synaptic terminals to suppress glutamate release [189–191]. It is suspected that glutamate acts post-

synaptically to trigger production of anandamide and other endogenous cannabinoid receptor agonists, which in turn act as a feedback signals to diminish pre-synaptic glutamate release. Cannabinoids and A2A antagonists both act presynaptically to decrease cAMP levels; cAMP up-regulates release of glutamate from glutamergic terminals [192–195]. The fact that CB1 knock-out mice are more susceptible to excitotoxicity [188] suggests that anandamide feedback is of physiological importance (and moreover raises some concern regarding the possible impact of longterm use of rimonabant – the CB1-antagonist appetite suppressant – on risk for neurodegenerative disorders). Cannabinoids protect PC12 pheochromocytoma cells from  $\beta$  amyloid toxicity, are protective in a transgenic mouse model of ALS, and limit infarct volume in focal cerebral ischemia [196–198]. However, in a rat model for Huntington's disease (striatal malonate injection), tetrahydrocannabinol had an adverse effects, possibly because of its agonist activity for CB2 receptors [199].

The fact that microglia can express CB2 receptors suggests that selective CB2 agonists – which are not psychoactive – could have some neuroprotective activity. However, the cannabinoid receptors expressed on neurons are exclusively CB1 – so selective CB2 agonists could not be expected to provide the same level of protection as would nonselective agonists such as tetrahydrocannabinol.

Synthetic cannabinoids are now being assessed clinically in traumatic brain injury and stroke. Centrally-acting cannabinoids should be evaluated in a wider range of rodent models of neurodegeneration. There does not yet appear to be any epidemiology focusing on risk for neurodegenerative disorders in habitual cannabis users; perhaps Jamaica would be an appropriate venue for such research. As a caution, it should be noted that marijuana use by young people has been linked to increase risk for schizophrenia [200].

## Sesamin

Various antioxidant phytonutrients, such as resveratrol, silymarin, and EGCG, have been shown to have a down-regulatory impact on microglial activation in vitro, presumably because these agents can inhibit NF- $\kappa$ B activation [201–204]. However, these effects require micromolar concentrations which would likely be impossible to sustain in vivo, owing to rapid metabolism of these agents. On the other hand, the intriguing lignan sesamin, a

prominent component of sesame seeds, not only inhibits the LPS-mediated activation of microglial cells in vitro [205,206], but also protects against rotenone-induced Parkinsonism when fed to rats [207]. This may reflect the fact that sesamin, lacking hydroxyl groups, is less readily susceptible to conjugation in the liver, and thus can achieve ample concentrations in the serum and the brain. Although this phytonutrient is not yet widely available as a supplement, it might have considerable potential for neuroprotection, and should be evaluated further.

## Complementary strategies

Suppression of microglial activation, by dampening excessive production of peroxynitrite and COX-2-derived prostanoids, can be expected to favorably impact the many neurodegenerative conditions in which excitotoxicity plays a prominent pathogenic role. However, there clearly are a number of additional strategies which might help to quell excitotoxicity – some of which would presumably be compatible with, and complementary to, microglial down-regulation.

## Improve astrocyte performance

Astrocytes protect neurons by sequestering extracellular glutamate, converting it to glutamine; the adverse impacts of peroxynitrite and of COX-2-derived prostanoids on this mechanism have been cited. Astrocytes also produce a range of neurotrophic factors which function to protect neurons from apoptosis and which can support the production of new neurons when appropriate [208–210]. As we have noted, calcitriol can increase astrocyte production of several neurotrophic factors: nerve growth factor, neurotrophin-3, and glial-derived growth factor. In rodents, caloric restriction, physical exercise, and environmental enrichment increase the expression of BDNF and of GDNF in various regions of the brain, including the cortex, hippocampus, and striatum [211–215]. The clinical correlate of this may be the reduced risk for Alzheimer's disease noted in those who engage in regular physical or mental exercise, or who consume relatively few calories [216–219]. Exercise training and lower-fat, lower-calorie diets also appear to be protective with respect to Parkinson's disease [220–223]. Mattson refers to caloric restriction, physical exercise, and mental activity as a type of "hormesis" – small stresses to the brain that up-regulate various

neuroprotective mechanisms which can help neurons to survive when faced with the larger stresses that trigger and sustain neurodegenerative disorders [211]. Other research is attempting to discover drugs which can act directly on astrocytes to promote production of neurotrophic factors; agents showing promise in this regard include various dopamine agonists, the anti-excitotoxic agent riluzole, the vasodilator ifenprodil, and the cognitive enhancer FK960 [224–228].

### Down-regulate presynaptic glutamate release

This mechanism may be crucial to the neuroprotective activity of cannabinoids and of A2A receptor inhibitors such as caffeine. Maintaining good neuron bioenergetics is also of importance in this regard; during strokes, neuron depolarization consequent to ischemia promotes glutamate release, triggering excitotoxicity. Thus, preserving efficient cerebral circulation (for example, with high-potassium, low-salt diets and hypertension control) should be of value in this regard, as should supplemental nutrients which support neuron bioenergetics (see below).

### Inhibit activation of NMDA receptors

Activated NMDA receptors are primarily responsible for excessive calcium influx during excitotoxicity. Unfortunately, most agents which directly inhibit NMDA receptors are unacceptably toxic, as they impair the long-term potentiation mechanism required for learning [229]. However, the drug memantine, long approved in Germany for the treatment of Alzheimer's disease, is a non-competitive NMDA receptor antagonist that has an intriguingly selective impact on these receptors – inhibiting the low-grade chronic activation of these receptors that is usually involved in excitotoxicity, while having relatively little impact on the sharp discrete signals required for long-term potentiation [230–232]. Thus, clinical tolerance to memantine is far superior to that for other NMDA antagonists. Memantine is effective in a range of rodent models of excitotoxicity, and is of documented clinical efficacy in Alzheimer's disease. Studies are underway to evaluate its utility in a wider range of neurodegenerative disorders. Another relatively well tolerated NMDA antagonist, the cough-suppressant dextromethorphan, may also have clinical potential as an anti-excitotoxic agent – albeit it did not improve survival in ALS in a recent clinical trial [233].

### Activate postsynaptic GABA(A) receptors

These receptors, expressed on many postsynaptic dendrites, act to hyperpolarize postsynaptic membranes by boosting chloride conductance. This in turn impedes calcium influx via activated NMDA receptors. During excitotoxic episodes, neurons release taurine and GABA into the extracellular space, where they can activate postsynaptic GABA(A) receptors, providing feedback suppression of excitotoxicity [234–238]. Since taurine supplementation can boost brain taurine stores – by about 50% in rats [239] – it seems likely that such supplementation could help to control excitotoxicity. While there is clear evidence that taurine can suppress excitotoxicity in vitro [237,240–242] – for example, inhibiting  $\beta$  amyloid-induced death in cultured chick retinal neurons [242] – the impact of supplemental taurine on ischemia-induced neuronal death in vivo is more equivocal. Thus, whereas taurine pre-administration has shown efficacy in some models of excitotoxicity [243,244], intracranial administration of taurine produced only modest non-significant protection in rat models of focal or global ischemia [245]. The extracellular taurine concentration measured in rat brains during ischemic episodes rises to the range of 5–18  $\mu\text{M}$  [234,235] – whereas the affinity of taurine for the GABA(A) receptor is said to be 40–50  $\mu\text{M}$  [238]. Thus, it is conceivable that the rise in extracellular taurine during excitotoxicity is of real, if perhaps modest, physiological significance as a feedback protective mechanism – in which case supplemental taurine would likely potentiate this neuroprotection. Taurine is reported to have modest and inconsistent efficacy for controlling epilepsy in children [246,247]; presumably, this could reflect a down-regulatory impact on glutamate transmission. Taurine also acts as a scavenger for the potent oxidant hypochlorous acid, which can be produced by activated microglia and possibly contributes to neurodegeneration [248]; moreover, taurine's natural derivative taurine chloramine can inhibit induction of iNOS in microglia [249].

The herb kava–kava, long used as a mild intoxicant by various South Pacific cultures, is a source of “kavapyrones” that appear to mediate its clinical activity in anxiety syndromes. These agents have been reported to induce a rapid up-regulation of GABA(A) receptor expression in the hippocampus and frontal cortex of rats [250]; however, they have no direct agonist activity for these receptors – possibly explaining why kava is better tolerated



and less habit-forming than direct agonists for these receptors such as benzodiazepines. The possibility that kava-kava may have neuroprotective activity is supported by a rat study demonstrating that kava extracts are approximately as effective as memantine in decreasing infarct volume following occlusion of the left middle cerebral artery [251]. Kava's utility in this regard – like that of memantine – may be more general, and it would be of interest to test it in the context of concurrent taurine supplementation.

### Boost neuronal protective mechanisms

Neurons express a range of proteins – antioxidant enzymes, heat shock proteins, calcium sequestrants, anti-apoptotic factors – which can protect them from the potentially lethal consequences of calcium overload. It may be feasible to up-regulate the expression of these protective proteins. In particular, the broad spectrum neuroprotective utility of caloric restriction or intermittent fasting in rodents and monkeys, may largely reflect increased neuronal expression of a range of protective proteins – as well as a decrease in expression of pro-apoptotic glucocorticoid receptors [211,252,253]. Some, but probably not all, of this effect is secondary to increase production of neurotrophic factors, which function to promote neuron survival. Regular exercise and mental stimulation also have some efficacy in this regard [211]. The antioxidant capacity of neurons can also be boosted with the nutrient lipoic acid; in sufficient concentrations, this acts as a phase II inducer, increasing neuronal synthesis of glutathione while also increasing the expression of various antioxidant enzymes [254–256]. Presumably, this explains the versatile neuroprotective activity of lipoic acid in rodents [257–263]. Lipoic acid is a well tolerated nutrient – though the oral doses required for optimal efficacy (600–1800 mg daily have shown some clinical efficacy in diabetic neuropathy) [264] can be expensive. Maintaining adequate selenium status – of potential importance in regions of the world where soil selenium is low – should also support the antioxidant defenses of neurons [265–268]. Although the impact of  $\alpha$ -tocopherol on neurodegenerative disorders is receiving attention – in light of evidence that lipid peroxides are mediators of the adverse impact of oxidative stress on neural function [32] – it would also be of interest to evaluate  $\gamma$ -tocopherol in this regard, inasmuch as this agent has a peroxynitrite-scavenging activity not possessed by  $\alpha$ -tocopherol [269,270].

### Support neuronal bioenergetics

Creatine acts as a “energy buffer” in excitable tissues that have rapidly varying energy requirements – such as muscles and neurons. Beal and colleagues have shown that supplemental creatine – which can boost brain stores of this nutrient – has versatile neuroprotective activity [271–274]. Other nutrients which appear to aid neuron bioenergetics under certain circumstances include coenzyme Q10 and acetylcarnitine [275–278]; a small clinical study has concluded that, at a dose of 1200 mg daily, coenzyme Q10 can slow clinical deterioration in Parkinson's disease [276]. Ketone bodies, which serve as alternative fuel for the CNS during fasting metabolism, can improve the bioenergetics of neurons when pyruvate dehydrogenase is suboptimally active; presumably, this is why ketotic diets are useful in the management of pediatric epilepsy, and why ketone infusion is protective in MPTP-induced neuropathy [279–281]. These findings point to the possible utility of medium-chain triglycerides (converted to ketone bodies in the liver) in the prevention and management of neurodegenerative syndromes [282].

Efficient cerebrovascular perfusion is evidently a *sine qua non* for optimal neuronal bioenergetics. Small strokes and diminished vascular perfusion are suspected to play a co-factor role in the induction of Alzheimer's dementia [283]. A low-salt, potassium-rich whole foods diet – the type of diet that humans evolved with – is associated with a low risk for hypertension and an even lower risk for stroke. On the Melanesian island of Kitava, whose inhabitants still do not salt their food, potassium intakes (primarily from yams) are as high as 8 g daily, the diet is quasi-vegan (small amounts of fish are consumed), and most people remain lean and insulin sensitive throughout life. Stroke appears to be rare or nonexistent among these people – many of whom live to an advanced age - and the very concept of senile dementia is unknown [284–286]. A similar rarity of senile dementia was reported among black East Africans during the early twentieth century, when salt use, hypertension, and stroke were still rare [286,287]. This suggests that preserving efficient cerebrovascular perfusion into old age may have a remarkably favorable impact on risk for dementia – not only vascular dementia, but also Alzheimer's. In Western epidemiology, obesity and hyperinsulinemia [288,289] – which may increase brain production of  $\beta$  amyloid [290] – have been linked to increased Alzheimer's risk, so the leanness of the Kitavans may contribute to their freedom from dementia;

nonetheless, stroke has been common in lean East Asian societies which heavily salt their food.

### Promote proper coupling of nNOS

When neuronal levels of either arginine or tetrahydrobiopterin are suboptimal, activation of nNOS by excitotoxic calcium influx induces production of superoxide as well as of NO – giving rise to peroxynitrite [36,37]. Grima and colleagues have shown that arginine can protect neurons from glutamate toxicity in vitro [38]. Furthermore, they note that glutamate stimulates astrocytes to transfer arginine to neurons – a physiological mechanism that may aid control of excitotoxicity in vivo. Surprisingly, aside from a single study demonstrating that arginine pre-loading decreases infarct volume after simulated stroke (the benefit being attributed to improved cerebral perfusion) [291], there seem to have been few if any attempts to evaluate the impact of supplemental arginine on excitotoxic syndromes in rodents. Furthermore, little if any attention has been devoted to the possibility that tetrahydrobiopterin – readily oxidized by peroxynitrite [292] – might be suboptimally available in neurons threatened with excitotoxicity [37]. In this regard, it is intriguing to note that 5-methyltetrahydrofolate somehow compensates for tetrahydrobiopterin deficiency in endothelial cells, thereby accounting for the beneficial impact of high-dose folate on dysfunctional endothelium (independent of its impact on homocysteine levels) [293–296]. Could high-dose folate likewise have favorable impact on nNOS activity when tetrahydrobiopterin availability is limiting? Supplemental folate is already recommended for neuroprotection, inasmuch as elevated homocysteine has been found to be a risk factor for Alzheimer's disease and stroke [297,298]; however, the doses required to promote tetrahydrobiopterin function are higher than those required for homocysteine control.

### Employ a range of complementary strategies

In the future, it seems likely that effective neuroprotection will be achieved, not by some single “magic bullet” drug, but rather by a mélange of well tolerated agents and lifestyle measures that address complementary aspects of the neurodegenerative process. As a demonstration of this principle, Beal and colleagues have recently shown that joint administration of creatine and a COX-2 inhibitor is much more effective than either agent alone in stemming loss of dopaminergic neurons in the MPTP model of Parkinsonism, and in preserving

motor neurons in a transgenic mouse model of ALS [299,300].

Although neurodegenerative disorders share certain common features – microglial activation associated with excitotoxic neuronal death and dysfunction – they differ in the “trigger” mechanisms that target neurodegeneration to specific neural pathways. (For example,  $\beta$  amyloid overproduction in Alzheimer's, mutant superoxide dismutase in some cases of hereditary ALS). Thus, effective management or prevention of a specific neurodegenerative disorder may require measures which address the trigger specific to that disorder. Measures which provide symptomatic support by compensating for the neuronal losses characteristic of that disorder (e.g. L-DOPA in Parkinsonism, acetylcholinesterase inhibitors in Alzheimer's) will also continue to play an important role in therapeutic protocols.

### What we can do now

Healthy people desiring to minimize their risk for neurodegenerative disorders could reasonably include the following in their daily supplement regimens (finances permitting!): vitamin D, soy isoflavones, creatine, selenium, coenzyme Q10, acetylcarnitine, lipoic acid, and taurine. Ingesting several strong cups of coffee daily (or taking a caffeine supplement) can also be recommended in this regard, along with regular physical and mental exercise, and moderation in calorie intake. For stroke prevention, frequent ingestion of potassium-rich whole foods, coupled with moderation in salt intake, is particularly advisable.

Presumably, it will be several more years before clinical trials enable us to judge the true merits of minocycline, pioglitazone, memantine, statins (or policosanol) and COX-2 inhibitors as clinical neuroprotective agents. However, since these drugs are usually well tolerated and reasonably safe (assuming that mini-dose aspirin is taken in conjunction with COX-2 inhibitors, and statin use is monitored), it may not be imprudent for patients in the early stages of neurodegenerative disorders to use these drugs, providing that they can find a cooperative physician. They might also be well advised to avail themselves of the nutrients and lifestyle measures cited in the preceding paragraph.

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## References

- [1] Liu B, Hong JS. Role of microglia in inflammation-mediated neurodegenerative diseases: mechanisms and strategies for therapeutic intervention. *J Pharmacol Exp Ther* 2003;304:1–7.
- [2] Mrak RE, Griffin WS. Glia and their cytokines in progression of neurodegeneration. *Neurobiol Aging* 2005;26:349–54.
- [3] Blasko I, Stampfer-Kountchev M, Robatscher P, Veerhuis R, Eikelenboom P, Grubeck-Loebenstein B. How chronic inflammation can affect the brain and support the development of Alzheimer's disease in old age: the role of microglia and astrocytes. *Aging Cell* 2004;3:169–76.
- [4] Vila M, Jackson-Lewis V, Guegan C, Wu DC, Teismann P, Choi DK, et al. The role of glial cells in Parkinson's disease. *Curr Opin Neurol* 2001;14:483–9.
- [5] Teismann P, Tieu K, Cohen O, Choi DK, Wu DC, Marks D, et al. Pathogenic role of glial cells in Parkinson's disease. *Mov Disord* 2003;18:121–9.
- [6] Sargsyan SA, Monk PN, Shaw PJ. Microglia as potential contributors to motor neuron injury in amyotrophic lateral sclerosis. *Glia* 2005.
- [7] Sapp E, Kegel KB, Aronin N, Hashikawa T, Uchiyama Y, Tohyama K, et al. Early and progressive accumulation of reactive microglia in the Huntington disease brain. *J Neuropathol Exp Neurol* 2001;60:161–72.
- [8] Aldskogius H. Regulation of microglia – potential new drug targets in the CNS. *Expert Opin Ther Targets* 2001;5:655–68.
- [9] Torreilles F, Salman-Tabcheh S, Guerin M, Torreilles J. Neurodegenerative disorders: the role of peroxynitrite. *Brain Res Brain Res Rev* 1999;30:153–63.
- [10] Kim WK, Ko KH. Potentiation of *N*-methyl-*D*-aspartate-mediated neurotoxicity by immunostimulated murine microglia. *J Neurosci Res* 1998;54:17–26.
- [11] Liberatore GT, Jackson-Lewis V, Vukosavic S, Mandir AS, Vila M, McAuliffe WG, et al. Inducible nitric oxide synthase stimulates dopaminergic neurodegeneration in the MPTP model of Parkinson disease. *Nat Med* 1999;5:1403–9.
- [12] Liu B, Gao HM, Wang JY, Jeohn GH, Cooper CL, Hong JS. Role of nitric oxide in inflammation-mediated neurodegeneration. *Ann NY Acad Sci* 2002;962:318–31.
- [13] Qin L, Liu Y, Cooper C, Liu B, Wilson B, Hong JS. Microglia enhance  $\beta$ -amyloid peptide-induced toxicity in cortical and mesencephalic neurons by producing reactive oxygen species. *J Neurochem* 2002;83:973–83.
- [14] Wu DC, Teismann P, Tieu K, Vila M, Jackson-Lewis V, Ischiropoulos H, et al. NADPH oxidase mediates oxidative stress in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine model of Parkinson's disease. *Proc Natl Acad Sci USA* 2003;100:6145–50.
- [15] Gao HM, Liu B, Hong JS. Critical role for microglial NADPH oxidase in rotenone-induced degeneration of dopaminergic neurons. *J Neurosci* 2003;23:6181–7.
- [16] Zekry D, Epperson TK, Krause KH. A role for NOX NADPH oxidases in Alzheimer's disease and other types of dementia?. *IUBMB Life* 2003;55:307–13.
- [17] Gao HM, Liu B, Zhang W, Hong JS. Critical role of microglial NADPH oxidase-derived free radicals in the in vitro MPTP model of Parkinson's disease. *FASEB J* 2003;17:1954–6.
- [18] Qin L, Liu Y, Wang T, Wei SJ, Block ML, Wilson B, et al. NADPH oxidase mediates lipopolysaccharide-induced neurotoxicity and proinflammatory gene expression in activated microglia. *J Biol Chem* 2004;279:1415–21.
- [19] Zhang W, Wang T, Qin L, Gao HM, Wilson B, Ali SF, et al. Neuroprotective effect of dextromethorphan in the MPTP Parkinson's disease model: role of NADPH oxidase. *FASEB J* 2004;18:589–91.
- [20] Zhao W, Xie W, Le W, Beers DR, He Y, Henkel JS, et al. Activated microglia initiate motor neuron injury by a nitric oxide and glutamate-mediated mechanism. *J Neuropathol Exp Neurol* 2004;63:964–77.
- [21] Beal MF. Excitotoxicity and nitric oxide in Parkinson's disease pathogenesis. *Ann Neurol* 1998;44:S110–4.
- [22] Doble A. The role of excitotoxicity in neurodegenerative disease: implications for therapy. *Pharmacol Ther* 1999;81:163–221.
- [23] Mattson MP. Excitotoxic and excitoprotective mechanisms: abundant targets for the prevention and treatment of neurodegenerative disorders. *Neuromolecular Med* 2003;3:65–94.
- [24] Arundine M, Tymianski M. Molecular mechanisms of calcium-dependent neurodegeneration in excitotoxicity. *Cell Calcium* 2003;34:325–37.
- [25] Mattson MP, Chan SL. Neuronal and glial calcium signaling in Alzheimer's disease. *Cell Calcium* 2003;34:385–97.
- [26] Novelli A, Reilly JA, Lysko PG, Henneberry RC. Glutamate becomes neurotoxic via the *N*-methyl-*D*-aspartate receptor when intracellular energy levels are reduced. *Brain Res* 1988;451:205–12.
- [27] Riobo NA, Clementi E, Melani M, Boveris A, Cadenas E, Moncada S, et al. Nitric oxide inhibits mitochondrial NADH: ubiquinone reductase activity through peroxynitrite formation. *Biochem J* 2001;359:139–45.
- [28] Radi R, Cassina A, Hodara R, Quijano C, Castro L. Peroxynitrite reactions and formation in mitochondria. *Free Radic Biol Med* 2002;33:1451–64.
- [29] Stewart VC, Heales SJ. Nitric oxide-induced mitochondrial dysfunction: implications for neurodegeneration. *Free Radic Biol Med* 2003;34:287–303.
- [30] Murray J, Taylor SW, Zhang B, Ghosh SS, Capaldi RA. Oxidative damage to mitochondrial complex I due to peroxynitrite: identification of reactive tyrosines by mass spectrometry. *J Biol Chem* 2003;278:37223–30.
- [31] Mark RJ, Hensley K, Butterfield DA, Mattson MP. Amyloid  $\beta$ -peptide impairs ion-motive ATPase activities: evidence for a role in loss of neuronal  $Ca^{2+}$  homeostasis and cell death. *J Neurosci* 1995;15:6239–49.
- [32] Mark RJ, Lovell MA, Markesbery WR, Uchida K, Mattson MP. A role for 4-hydroxynonenal, an aldehydic product of lipid peroxidation, in disruption of ion homeostasis and neuronal death induced by amyloid  $\beta$ -peptide. *J Neurochem* 1997;68:255–64.
- [33] Trotti D, Rossi D, Gjesdal O, Levy LM, Racagni G, Danbolt NC, et al. Peroxynitrite inhibits glutamate transporter subtypes. *J Biol Chem* 1996;271:5976–9.
- [34] Sorg O, Horn TF, Yu N, Gruol DL, Bloom FE. Inhibition of astrocyte glutamate uptake by reactive oxygen species: role of antioxidant enzymes. *Mol Med* 1997;3:431–40.
- [35] Ayata C, Ayata G, Hara H, Matthews RT, Beal MF, Ferrante RJ, et al. Mechanisms of reduced striatal NMDA excitotoxicity in type I nitric oxide synthase knock-out mice. *J Neurosci* 1997;17:6908–17.
- [36] Xia Y, Dawson VL, Dawson TM, Snyder SH, Zweier JL. Nitric oxide synthase generates superoxide and nitric oxide in arginine-depleted cells leading to peroxynitrite-mediated cellular injury. *Proc Natl Acad Sci USA* 1996;93:6770–4.

- [37] Vasquez-Vivar J, Hogg N, Martasek P, Karoui H, Pritchard Jr KA, Kalyanaram B. Tetrahydrobiopterin-dependent inhibition of superoxide generation from neuronal nitric oxide synthase. *J Biol Chem* 1999;274:26736–42.
- [38] Grima G, Benz B, Do KQ. Glial-derived arginine, the nitric oxide precursor, protects neurons from NMDA-induced excitotoxicity. *Eur J Neurosci* 2001;14:1762–70.
- [39] Huang Z, Huang PL, Panahian N, Dalkara T, Fishman MC, Moskowitz MA. Effects of cerebral ischemia in mice deficient in neuronal nitric oxide synthase. *Science* 1994;265:1883–5.
- [40] Dawson VL, Kizushi VM, Huang PL, Snyder SH, Dawson TM. Resistance to neurotoxicity in cortical cultures from neuronal nitric oxide synthase-deficient mice. *J Neurosci* 1996;16:2479–87.
- [41] Bauer MK, Lieb K, Schulze-Osthoff K, Berger M, Gebicke-Haerter PJ, Bauer J, et al. Expression and regulation of cyclooxygenase-2 in rat microglia. *Eur J Biochem* 1997;243:726–31.
- [42] Akundi RS, Candelario-Jalil E, Hess S, Hull M, Lieb K, Gebicke-Haerter PJ, et al. Signal transduction pathways regulating cyclooxygenase-2 in lipopolysaccharide-activated primary rat microglia. *Glia* 2005.
- [43] Bezzi P, Carmignoto G, Pasti L, Vesce S, Rossi D, Rizzini BL, et al. Prostaglandins stimulate calcium-dependent glutamate release in astrocytes. *Nature* 1998;391:281–5.
- [44] Casper D, Yaparalvi U, Rempel N, Werner P. Ibuprofen protects dopaminergic neurons against glutamate toxicity in vitro. *Neurosci Lett* 2000;289:201–4.
- [45] Hewett SJ, Uliasz TF, Vidwans AS, Hewett JA. Cyclooxygenase-2 contributes to N-methyl-D-aspartate-mediated neuronal cell death in primary cortical cell culture. *J Pharmacol Exp Ther* 2000;293:417–25.
- [46] Salzberg-Brenhouse HC, Chen EY, Emerich DF, Baldwin S, Hogeland K, Ranelli S, et al. Inhibitors of cyclooxygenase-2, but not cyclooxygenase-1 provide structural and functional protection against quinolinic acid-induced neurodegeneration. *J Pharmacol Exp Ther* 2003;306:218–28.
- [47] Mirjany M, Ho L, Pasinetti GM. Role of cyclooxygenase-2 in neuronal cell cycle activity and glutamate-mediated excitotoxicity. *J Pharmacol Exp Ther* 2002;301:494–500.
- [48] Floden AM, Li S, Combs CK.  $\beta$ -Amyloid-stimulated microglia induce neuron death via synergistic stimulation of tumor necrosis factor  $\alpha$  and NMDA receptors. *J Neurosci* 2005;25:2566–75.
- [49] Glazner GW, Mattson MP. Differential effects of BDNF, ADNF9, and TNF $\alpha$  on levels of NMDA receptor subunits, calcium homeostasis, and neuronal vulnerability to excitotoxicity. *Exp Neurol* 2000;161:442–52.
- [50] Huang TL, O'Banion MK. Interleukin-1 $\beta$  and tumor necrosis factor- $\alpha$  suppress dexamethasone induction of glutamine synthetase in primary mouse astrocytes. *J Neurochem* 1998;71:1436–42.
- [51] Hu S, Sheng WS, Ehrlich LC, Peterson PK, Chao CC. Cytokine effects on glutamate uptake by human astrocytes. *Neuroimmunomodulation* 2000;7:153–9.
- [52] Liao SL, Chen CJ. Differential effects of cytokines and redox potential on glutamate uptake in rat cortical glial cultures. *Neurosci Lett* 2001;299:113–6.
- [53] Wang Z, Pekarskaya O, Bencheikh M, Chao W, Gelbard HA, Ghorpade A, et al. Reduced expression of glutamate transporter EAAT2 and impaired glutamate transport in human primary astrocytes exposed to HIV-1 or gp120. *Virology* 2003;312:60–73.
- [54] Rozovsky I, Finch CE, Morgan TE. Age-related activation of microglia and astrocytes: in vitro studies show persistent phenotypes of aging, increased proliferation, and resistance to down-regulation. *Neurobiol Aging* 1998;19:97–103.
- [55] Mackenzie IR, Munoz DG. Non-steroidal anti-inflammatory drug use and Alzheimer-type pathology in aging. *Neurology* 1998;50:986–90.
- [56] Sheng JG, Mrak RE, Griffin WS. Enlarged and phagocytic, but not primed, interleukin-1  $\alpha$ -immunoreactive microglia increase with age in normal human brain. *Acta Neuropathol (Berl)* 1998;95:229–34.
- [57] Barcia C, Sanchez BA, Fernandez-Villalba E, Bautista V, Poza YP, Fernandez-Barreiro A, et al. Evidence of active microglia in substantia nigra pars compacta of parkinsonian monkeys 1 year after MPTP exposure. *Glia* 2004;46:402–9.
- [58] Langston JW, Forno LS, Tetrad J, Reeves AG, Kaplan JA, Karluk D. Evidence of active nerve cell degeneration in the substantia nigra of humans years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine exposure. *Ann Neurol* 1999;46:598–605.
- [59] Hirsch EC, Breidert T, Rousset E, Hunot S, Hartmann A, Michel PP. The role of glial reaction and inflammation in Parkinson's disease. *Ann NY Acad Sci* 2003;991:214–28.
- [60] Jenner P. Oxidative mechanisms in nigral cell death in Parkinson's disease. *Mov Disord* 1998;13(Suppl. 1):24–34.
- [61] Gao HM, Jiang J, Wilson B, Zhang W, Hong JS, Liu B. Microglial activation-mediated delayed and progressive degeneration of rat nigral dopaminergic neurons: relevance to Parkinson's disease. *J Neurochem* 2002;81:1285–97.
- [62] Arimoto T, Bing G. Up-regulation of inducible nitric oxide synthase in the substantia nigra by lipopolysaccharide causes microglial activation and neurodegeneration. *Neurobiol Dis* 2003;12:35–45.
- [63] Choi SH, Joe EH, Kim SU, Jin BK. Thrombin-induced microglial activation produces degeneration of nigral dopaminergic neurons in vivo. *J Neurosci* 2003;23:5877–86.
- [64] Iravani MM, Kashefi K, Mander P, Rose S, Jenner P. Involvement of inducible nitric oxide synthase in inflammation-induced dopaminergic neurodegeneration. *Neuroscience* 2002;110:49–58.
- [65] Versijpt JJ, Dumont F, Van Laere KJ, Decoo D, Santens P, Audenaert K, et al. Assessment of neuroinflammation and microglial activation in Alzheimer's disease with radiolabelled PK11195 and single photon emission computed tomography. A pilot study. *Eur Neurol* 2003;50:39–47.
- [66] Bianca VD, Dusi S, Bianchini E, Dal PI, Rossi F.  $\beta$ -Amyloid activates the O-2 forming NADPH oxidase in microglia, monocytes, and neutrophils. A possible inflammatory mechanism of neuronal damage in Alzheimer's disease. *J Biol Chem* 1999;274:15493–9.
- [67] Ge YW, Lahiri DK. Regulation of promoter activity of the APP gene by cytokines and growth factors: implications in Alzheimer's disease. *Ann NY Acad Sci* 2002;973:463–7.
- [68] Blasko I, Marx F, Steiner E, Hartmann T, Grubeck-Loebenstein B. TNF $\alpha$  plus IFN $\gamma$  induce the production of Alzheimer  $\beta$ -amyloid peptides and decrease the secretion of APPs. *FASEB J* 1999;13:63–8.
- [69] Sastre M, Dewachter I, Landreth GE, Willson TM, Klockgether T, Van Leuven F, et al. Non-steroidal anti-inflammatory drugs and peroxisome proliferator-activated receptor- $\gamma$  agonists modulate immunostimulated processing of amyloid precursor protein through regulation of  $\beta$ -secretase. *J Neurosci* 2003;23:9796–804.
- [70] Qin W, Ho L, Pompl PN, Peng Y, Zhao Z, Xiang Z, et al. Cyclooxygenase (COX)-2 and COX-1 potentiate  $\beta$ -amyloid

- peptide generation through mechanisms that involve  $\gamma$ -secretase activity. *J Biol Chem* 2003;278:50970–7.
- [71] Tikka T, Fiebich BL, Goldsteins G, Keinanen R, Koistinaho J. Minocycline, a tetracycline derivative, is neuroprotective against excitotoxicity by inhibiting activation and proliferation of microglia. *J Neurosci* 2001;21:2580–8.
- [72] He Y, Appel S, Le W. Minocycline inhibits microglial activation and protects nigral cells after 6-hydroxydopamine injection into mouse striatum. *Brain Res* 2001;909:187–93.
- [73] Kim SS, Kong PJ, Kim BS, Sheen DH, Nam SY, Chun W. Inhibitory action of minocycline on lipopolysaccharide-induced release of nitric oxide and prostaglandin E2 in BV2 microglial cells. *Arch Pharm Res* 2004;27:314–8.
- [74] Suk K. Minocycline suppresses hypoxic activation of rodent microglia in culture. *Neurosci Lett* 2004;366:167–71.
- [75] Blum D, Chtarto A, Tenenbaum L, Brotchi J, Levivier M. Clinical potential of minocycline for neurodegenerative disorders. *Neurobiol Dis* 2004;17:359–66.
- [76] Zemke D, Majid A. The potential of minocycline for neuroprotection in human neurologic disease. *Clin Neuropharmacol* 2004;27:293–8.
- [77] Pocock JM, Liddle AC. Microglial signalling cascades in neurodegenerative disease. *Prog Brain Res* 2001;132:555–65.
- [78] Bernardo A, Levi G, Minghetti L. Role of the peroxisome proliferator-activated receptor- $\gamma$  (PPAR- $\gamma$ ) and its natural ligand 15-deoxy- $\Delta$ 12,14-prostaglandin J2 in the regulation of microglial functions. *Eur J Neurosci* 2000;12:2215–23.
- [79] Kim EJ, Kwon KJ, Park JY, Lee SH, Moon CH, Baik EJ. Effects of peroxisome proliferator-activated receptor agonists on LPS-induced neuronal death in mixed cortical neurons: associated with iNOS and COX-2. *Brain Res* 2002;941:1–10.
- [80] Storer PD, Xu J, Chavis JA, Drew PD. Cyclopentenone prostaglandins PGA(2) and 15-deoxy- $\delta$ (12,14) PGJ(2) suppress activation of murine microglia and astrocytes: implications for multiple sclerosis. *J Neurosci Res* 2005;80:66–74.
- [81] Dehmer T, Heneka MT, Sastre M, Dichgans J, Schulz JB. Protection by pioglitazone in the MPTP model of Parkinson's disease correlates with  $\kappa$  B  $\alpha$  induction and block of NF  $\kappa$  B and iNOS activation. *J Neurochem* 2004;88:494–501.
- [82] Breidert T, Callebert J, Heneka MT, Landreth G, Launay JM, Hirsch EC. Protective action of the peroxisome proliferator-activated receptor- $\gamma$  agonist pioglitazone in a mouse model of Parkinson's disease. *J Neurochem* 2002;82:615–24.
- [83] Camacho IE, Serneels L, Spittaels K, Merchiers P, Dominguez D, De Strooper B. Peroxisome proliferator-activated receptor  $\gamma$  induces a clearance mechanism for the amyloid- $\beta$  peptide. *J Neurosci* 2004;24:10908–17.
- [84] Inestrosa NC, Godoy JA, Quintanilla RA, Koenig CS, Bronfman M. Peroxisome proliferator-activated receptor  $\gamma$  is expressed in hippocampal neurons and its activation prevents  $\beta$ -amyloid neurodegeneration: role of Wnt signaling. *Exp Cell Res* 2005;304:91–104.
- [85] Feinstein DL, Galea E, Gavrilyuk V, Brosnan CF, Whitacre CC, Dumitrescu-Ozimek L, et al. Peroxisome proliferator-activated receptor- $\gamma$  agonists prevent experimental autoimmune encephalomyelitis. *Ann Neurol* 2002;51:694–702.
- [86] Pershadsingh HA, Heneka MT, Saini R, Amin NM, Broeske DJ, Feinstein DL. Effect of pioglitazone treatment in a patient with secondary multiple sclerosis. *J Neuroinflammation* 2004;1:3.
- [87] Uryu S, Harada J, Hisamoto M, Oda T. Troglitazone inhibits both post-glutamate neurotoxicity and low-potassium-induced apoptosis in cerebellar granule neurons. *Brain Res* 2002;924:229–36.
- [88] Benedetti MD, Maraganore DM, Bower JH, McDonnell SK, Peterson BJ, Ahlskog JE, et al. Hysterectomy, menopause, and estrogen use preceding Parkinson's disease: an exploratory case-control study. *Mov Disord* 2001;16:830–7.
- [89] Sawada H, Shimohama S. Estrogens and Parkinson disease: novel approach for neuroprotection. *Endocrine* 2003;21:77–9.
- [90] Wooten GF, Currie LJ, Bovbjerg VE, Lee JK, Patrie J. Are men at greater risk for Parkinson's disease than women?. *J Neurol Neurosurg Psychiatry* 2004;75:637–9.
- [91] Currie LJ, Harrison MB, Trugman JM, Bennett JP, Wooten GF. Postmenopausal estrogen use affects risk for Parkinson disease. *Arch Neurol* 2004;61:886–8.
- [92] Vegeto E, Bonincontro C, Pollio G, Sala A, Viappiani S, Nardi F, et al. Estrogen prevents the lipopolysaccharide-induced inflammatory response in microglia. *J Neurosci* 2001;21:1809–18.
- [93] Baker AE, Brautigam VM, Watters JJ. Estrogen modulates microglial inflammatory mediator production via interactions with estrogen receptor  $\beta$ . *Endocrinology* 2004;145:5021–32.
- [94] Dluzen DE, McDermott JL, Liu B. Estrogen alters MPTP-induced neurotoxicity in female mice: effects on striatal dopamine concentrations and release. *J Neurochem* 1996;66:658–66.
- [95] Grandbois M, Morissette M, Callier S, Di Paolo T. Ovarian steroids and raloxifene prevent MPTP-induced dopamine depletion in mice. *Neuroreport* 2000;11:343–6.
- [96] Ramirez AD, Liu X, Menniti FS. Repeated estradiol treatment prevents MPTP-induced dopamine depletion in male mice. *Neuroendocrinology* 2003;77:223–31.
- [97] D'Astous M, Morissette M, Di Paolo T. Effect of estrogen receptor agonists treatment in MPTP mice: evidence of neuroprotection by an ER  $\alpha$  agonist. *Neuropharmacology* 2004;47:1180–8.
- [98] Henderson VW. Hormone therapy and Alzheimer's disease: benefit or harm? *Expert Opin Pharmacother* 2004;5:389–406.
- [99] Brinton RD. Impact of estrogen therapy on Alzheimer's disease: a fork in the road?. *CNS Drugs* 2004;18:405–22.
- [100] Wang L, Andersson S, Warner M, Gustafsson JA. Morphological abnormalities in the brains of estrogen receptor  $\beta$  knockout mice. *Proc Natl Acad Sci USA* 2001;98:2792–6.
- [101] Takahashi N, Tonchev AB, Koike K, Murakami K, Yamada K, Yamashita T, et al. Expression of estrogen receptor- $\beta$  in the postischemic monkey hippocampus. *Neurosci Lett* 2004;369:9–13.
- [102] Savaskan E, Olivieri G, Meier F, Ravid R, Muller-Spahn F. Hippocampal estrogen  $\beta$ -receptor immunoreactivity is increased in Alzheimer's disease. *Brain Res* 2001;908:113–9.
- [103] Trieu VN, Uckun FM. Genistein is neuroprotective in murine models of familial amyotrophic lateral sclerosis and stroke. *Biochem Biophys Res Commun* 1999;258:685–8.
- [104] Bang OY, Hong HS, Kim DH, Kim H, Boo JH, Huh K, et al. Neuroprotective effect of genistein against  $\beta$ -amyloid-induced neurotoxicity. *Neurobiol Dis* 2004;16:21–8.
- [105] Zeng H, Chen Q, Zhao B. Genistein ameliorates  $\beta$ -amyloid peptide (25–35)-induced hippocampal neuronal apoptosis. *Free Radic Biol Med* 2004;36:180–8.

- [106] Wang X, Chen S, Ma G, Ye M, Lu G. Genistein protects dopaminergic neurons by inhibiting microglial activation. *Neuroreport* 2005;16:267–70.
- [107] McCarty MF. Isoflavones made simple – genistein’s agonist activity for the  $\beta$ -type estrogen receptor mediates their health benefits. *Med Hypotheses* 2006;66:1093–114.
- [108] Squadrito F, Altavilla D, Morabito N, Crisafulli A, D’Anna R, Corrado F, et al. The effect of the phytoestrogen genistein on plasma nitric oxide concentrations, endothelin-1 levels and endothelium dependent vasodilation in postmenopausal women. *Atherosclerosis* 2002;163:339–47.
- [109] Morabito N, Crisafulli A, Vergara C, Gaudio A, Lasco A, Frisina N, et al. Effects of genistein and hormone-replacement therapy on bone loss in early postmenopausal women: a randomized double-blind placebo-controlled study. *J Bone Miner Res* 2002;17:1904–12.
- [110] Dubal DB, Shughrue PJ, Wilson ME, Merchenthaler I, Wise PM. Estradiol modulates bcl-2 in cerebral ischemia: a potential role for estrogen receptors. *J Neurosci* 1999;19:6385–93.
- [111] Zhao L, Wu TW, Brinton RD. Estrogen receptor subtypes  $\alpha$  and  $\beta$  contribute to neuroprotection and increased Bcl-2 expression in primary hippocampal neurons. *Brain Res* 2004;1010:22–34.
- [112] Carswell HV, Macrae IM, Gallagher L, Harrop E, Horsburgh KJ. Neuroprotection by a selective estrogen receptor  $\beta$  agonist in a mouse model of global ischemia. *Am J Physiol Heart Circ Physiol* 2004;287:H1501–4.
- [113] Miller NR, Jover T, Cohen HW, Zukin RS, Etgen AM. Estrogen can act via estrogen receptor  $\alpha$  and  $\beta$  to protect hippocampal neurons against global ischemia-induced cell death. *Endocrinology* 2005.
- [114] Sawada H, Ibi M, Kihara T, Honda K, Nakamizo T, Kanki R, et al. Estradiol protects dopaminergic neurons in a MPP+Parkinson’s disease model. *Neuropharmacology* 2002;42:1056–64.
- [115] Garcion E, Sindji L, Montero-Menei C, Andre C, Brachet P, Darcy F. Expression of inducible nitric oxide synthase during rat brain inflammation: regulation by 1,25-dihydroxyvitamin D<sub>3</sub>. *Glia* 1998;22:282–94.
- [116] Garcion E, Wion-Barbot N, Montero-Menei CN, Berger F, Wion D. New clues about vitamin D functions in the nervous system. *Trends Endocrinol Metab* 2002;13:100–5.
- [117] Lefebvre DC, Montero-Menei CN, Bernard R, Couez D. Vitamin D<sub>3</sub> inhibits proinflammatory cytokines and nitric oxide production by the EOC13 microglial cell line. *J Neurosci Res* 2003;71:575–82.
- [118] Naveilhan P, Neveu I, Wion D, Brachet P. 1,25-Dihydroxyvitamin D<sub>3</sub>, an inducer of glial cell line-derived neurotrophic factor. *Neuroreport* 1996;7:2171–5.
- [119] Sanchez B, Lopez-Martin E, Segura C, Labandeira-Garcia JL, Perez-Fernandez R. 1,25-Dihydroxyvitamin D(3) increases striatal GDNF mRNA and protein expression in adult rats. *Brain Res Mol Brain Res* 2002;108:143–6.
- [120] Wang JY, Wu JN, Cherng TL, Hoffer BJ, Chen HH, Borlongan CV, et al. Vitamin D(3) attenuates 6-hydroxydopamine-induced neurotoxicity in rats. *Brain Res* 2001;904:67–75.
- [121] Garcion E, Nataf S, Berod A, Darcy F, Brachet P. 1,25-Dihydroxyvitamin D<sub>3</sub> inhibits the expression of inducible nitric oxide synthase in rat central nervous system during experimental allergic encephalomyelitis. *Brain Res Mol Brain Res* 1997;45:255–67.
- [122] Neveu I, Naveilhan P, Menaa C, Wion D, Brachet P, Garabedian M. Synthesis of 1,25-dihydroxyvitamin D<sub>3</sub> by rat brain macrophages in vitro. *J Neurosci Res* 1994;38:214–20.
- [123] Treves TA, Pedro-Cuesta J. Parkinsonism mortality in the US. 1. Time and space distribution. *Acta Neurol Scand* 1991;84:389–97.
- [124] Eisen A, Calne D. Amyotrophic lateral sclerosis, Parkinson’s disease and Alzheimer’s disease: phylogenetic disorders of the human neocortex sharing many characteristics. *Can J Neurol Sci* 1992;19:117–23.
- [125] Betemps EJ, Buncher CR. Birthplace as a risk factor in motor neurone disease and Parkinson’s disease. *Int J Epidemiol* 1993;22:898–904.
- [126] Vieth R. Vitamin D supplementation, 25-hydroxyvitamin D concentrations, and safety. *Am J Clin Nutr* 1999;69:842–56 [JID – 0376027].
- [127] Chen H, Zhang SM, Hernan MA, Schwarzschild MA, Willett WC, Colditz GA, et al. Non-steroidal anti-inflammatory drugs and the risk of Parkinson disease. *Arch Neurol* 2003;60:1059–64.
- [128] McGeer PL, Schulzer M, McGeer EG. Arthritis and anti-inflammatory agents as possible protective factors for Alzheimer’s disease: a review of 17 epidemiologic studies. *Neurology* 1996;47:425–32.
- [129] Minghetti L, Nicolini A, Polazzi E, Creminon C, Macclouf J, Levi G. Inducible nitric oxide synthase expression in activated rat microglial cultures is downregulated by exogenous prostaglandin E<sub>2</sub> and by cyclooxygenase inhibitors. *Glia* 1997;19:152–60.
- [130] Klegeris A, McGeer PL. Cyclooxygenase and 5-lipoxygenase inhibitors protect against mononuclear phagocyte neurotoxicity. *Neurobiol Aging* 2002;23:787–94.
- [131] Teismann P, Ferger B. Inhibition of the cyclooxygenase isoenzymes COX-1 and COX-2 provide neuroprotection in the MPTP-mouse model of Parkinson’s disease. *Synapse* 2001;39:167–74.
- [132] Yan Q, Zhang J, Liu H, Babu-Khan S, Vassar R, Biere AL, et al. Anti-inflammatory drug therapy alters  $\beta$ -amyloid processing and deposition in an animal model of Alzheimer’s disease. *J Neurosci* 2003;23:7504–9.
- [133] Drachman DB, Rothstein JD. Inhibition of cyclooxygenase-2 protects motor neurons in an organotypic model of amyotrophic lateral sclerosis. *Ann Neurol* 2000;48:792–5.
- [134] Levesque LE, Brophy JM, Zhang B. The risk for myocardial infarction with cyclooxygenase-2 inhibitors: a population study of elderly adults. *Ann Intern Med* 2005.
- [135] Kalmijn S, Launer LJ, Ott A, Witteman JC, Hofman A, Breteler MM. Dietary fat intake and the risk of incident dementia in the Rotterdam study. *Ann Neurol* 1997;42:776–82.
- [136] Grant WB, Campbell A, Itzhaki RF, Savory J. The significance of environmental factors in the etiology of Alzheimer’s disease. *J Alzheimers Dis* 2002;4:179–89.
- [137] Morris MC, Evans DA, Bienias JL, Tangney CC, Bennett DA, Wilson RS, et al. Consumption of fish and *n*-3 fatty acids and risk of incident Alzheimer disease. *Arch Neurol* 2003;60:940–6.
- [138] Li G, Higdon R, Kukull WA, Peskind E, Van Valen MK, Tsuang D, et al. Statin therapy and risk of dementia in the elderly: a community-based prospective cohort study. *Neurology* 2004;63:1624–8.
- [139] Zandi PP, Sparks DL, Khachaturian AS, Tschanz J, Norton M, Steinberg M, et al. Do statins reduce risk of incident dementia and Alzheimer disease? The Cache county study. *Arch Gen Psychiatry* 2005;62:217–24.
- [140] Wolozin B, Kellman W, Ruosseau P, Celesia GG, Siegel G. Decreased prevalence of Alzheimer disease associated with 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors. *Arch Neurol* 2000;57:1439–43.

- [141] Jick H, Zornberg GL, Jick SS, Seshadri S, Drachman DA. Statins and the risk of dementia. *Lancet* 2000;356:1627–31.
- [142] Rockwood K, Kirkland S, Hogan DB, MacKnight C, Merry H, Verreault R, et al. Use of lipid-lowering agents, indication bias, and the risk of dementia in community-dwelling elderly people. *Arch Neurol* 2002;59:223–7.
- [143] Zamrini E, McGwin G, Roseman JM. Association between statin use and Alzheimer's disease. *Neuroepidemiology* 2004;23:94–8.
- [144] Kuo YM, Emmerling MR, Bisgaier CL, Essenburg AD, Lampert HC, Drumm D, et al. Elevated low-density lipoprotein in Alzheimer's disease correlates with brain abeta 1-42 levels. *Biochem Biophys Res Commun* 1998;252:711–5.
- [145] Lesser G, Kandiah K, Libow LS, Likourezos A, Breuer B, Marin D, et al. Elevated serum total and LDL cholesterol in very old patients with Alzheimer's disease. *Dement Geriatr Cogn Disord* 2001;12:138–45.
- [146] Suryadevara V, Storey SG, Aronow WS, Ahn C. Association of abnormal serum lipids in elderly persons with atherosclerotic vascular disease and dementia, atherosclerotic vascular disease without dementia, dementia without atherosclerotic vascular disease, and no dementia or atherosclerotic vascular disease. *J Gerontol A Biol Sci Med Sci* 2003;58:M859–61.
- [147] Sabbagh M, Zahiri HR, Ceimo J, Cooper K, Gaul W, Connor D, et al. Is there a characteristic lipid profile in Alzheimer's disease?. *J Alzheimers Dis* 2004;6:585–9.
- [148] Lieberman A, Lyons K, Levine J, Myerburg R. Statins, cholesterol, co-enzyme Q10, and Parkinson's disease. *Parkinsonism Relat Disord* 2005;11:81–4.
- [149] Cordle A, Landreth G. 3-Hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors attenuate  $\beta$ -amyloid-induced microglial inflammatory responses. *J Neurosci* 2005;25:299–307.
- [150] Pahan K, Sheikh FG, Namboodiri AM, Singh I. Lovastatin and phenylacetate inhibit the induction of nitric oxide synthase and cytokines in rat primary astrocytes, microglia, and macrophages. *J Clin Invest* 1997;100:2671–9.
- [151] Naidu A, Xu Q, Catalano R, Cordell B. Secretion of apolipoprotein E by brain glia requires protein prenylation and is suppressed by statins. *Brain Res* 2002;958:100–11.
- [152] Bi X, Baudry M, Liu J, Yao Y, Fu L, Brucher F, et al. Inhibition of geranylgeranylation mediates the effects of 3-hydroxy-3-methylglutaryl (HMG)-CoA reductase inhibitors on microglia. *J Biol Chem* 2004;279:48238–45.
- [153] Fassbender K, Simons M, Bergmann C, Stroick M, Lutjohann D, Keller P, et al. Simvastatin strongly reduces levels of Alzheimer's disease  $\beta$ -amyloid peptides Abeta 42 and Abeta 40 in vitro and in vivo. *Proc Natl Acad Sci USA* 2001;98:5856–61.
- [154] Parvathy S, Ehrlich M, Pedrini S, Diaz N, Refolo L, Buxbaum JD, et al. Atorvastatin-induced activation of Alzheimer's  $\alpha$  secretase is resistant to standard inhibitors of protein phosphorylation-regulated ectodomain shedding. *J Neurochem* 2004;90:1005–10.
- [155] Cole SL, Grudzien A, Manhart IO, Kelly BL, Oakley H, Vassar R. Statins cause intracellular accumulation of amyloid precursor protein,  $\beta$ -secretase-cleaved fragments, and amyloid  $\beta$ -peptide via an isoprenoid-dependent mechanism. *J Biol Chem* 2005;280:18755–70.
- [156] Zacco A, Togo J, Spence K, Ellis A, Lloyd D, Furlong S, et al. 3-Hydroxy-3-methylglutaryl coenzyme A reductase inhibitors protect cortical neurons from excitotoxicity. *J Neurosci* 2003;23:11104–11.
- [157] Bosel J, Gandor F, Harms C, Synowitz M, Harms U, Djoufack PC, et al. Neuroprotective effects of atorvastatin against glutamate-induced excitotoxicity in primary cortical neurones. *J Neurochem* 2005;92:1386–98.
- [158] Sparks DL, Connor DJ, Browne PJ, Lopez JE, Sabbagh MN. HMG-CoA reductase inhibitors (statins) in the treatment of Alzheimer's disease and why it would be ill-advise to use one that crosses the blood-brain barrier. *J Nutr Health Aging* 2002;6:324–31.
- [159] Laufs U, La FV, Plutzky J, Liao JK. Upregulation of endothelial nitric oxide synthase by HMG CoA reductase inhibitors. *Circulation* 1998;97:1129–35.
- [160] Endres M, Laufs U, Huang Z, Nakamura T, Huang P, Moskowitz MA, et al. Stroke protection by 3-hydroxy-3-methylglutaryl (HMG)-CoA reductase inhibitors mediated by endothelial nitric oxide synthase. *Proc Natl Acad Sci USA* 1998;95:8880–5.
- [161] Jung KH, Chu K, Jeong SW, Han SY, Lee ST, Kim JY, et al. HMG-CoA reductase inhibitor, atorvastatin, promotes sensorimotor recovery, suppressing acute inflammatory reaction after experimental intracerebral hemorrhage. *Stroke* 2004;35:1744–9.
- [162] Sironi L, Cimino M, Guerrini U, Calvio AM, Lodetti B, Asdente M, et al. Treatment with statins after induction of focal ischemia in rats reduces the extent of brain damage. *Arterioscler Thromb Vasc Biol* 2003;23:322–7.
- [163] Menendez R, Amor AM, Rodeiro I, Gonzalez RM, Gonzalez PC, Alfonso JL, et al. Policosanol modulates HMG-CoA reductase activity in cultured fibroblasts. *Arch Med Res* 2001;32:8–12.
- [164] Gouni-Berthold I, Berthold HK. Policosanol: clinical pharmacology and therapeutic significance of a new lipid-lowering agent. *Am Heart J* 2002;143:356–65.
- [165] Varady KA, Wang Y, Jones PJ. Role of policosanols in the prevention and treatment of cardiovascular disease. *Nutr Rev* 2003;61:376–83.
- [166] Noa M, Mas R, Mendoza S, Gamez R, Mendoza N, Gonzalez J. Policosanol prevents bone loss in ovariectomized rats. *Drugs Exp Clin Res* 2004;30:117–23.
- [167] Borg J. The neurotrophic factor, *n*-hexacosanol, reduces the neuronal damage induced by the neurotoxin, kainic acid. *J Neurosci Res* 1991;29:62–7.
- [168] McCarty MF. Policosanol safely down-regulates HMG-CoA reductase – potential as a component of the Esselstyn regimen. *Med Hypotheses* 2002;59:268–79.
- [169] Ascherio A, Zhang SM, Hernan MA, Kawachi I, Colditz GA, Speizer FE, et al. Prospective study of caffeine consumption and risk of Parkinson's disease in men and women. *Ann Neurol* 2001;50:56–63.
- [170] Maia L, de Mendonca A. Does caffeine intake protect from Alzheimer's disease?. *Eur J Neurol* 2002;9:377–82.
- [171] Lindsay J, Laurin D, Verreault R, Hebert R, Helliwell B, Hill GB, et al. Risk factors for Alzheimer's disease: a prospective analysis from the Canadian study of health and aging. *Am J Epidemiol* 2002;156:445–53.
- [172] Schwarzschild MA, Xu K, Oztas E, Petzer JP, Castagnoli K, Castagnoli Jr N, et al. Neuroprotection by caffeine and more specific A2A receptor antagonists in animal models of Parkinson's disease. *Neurology* 2003;61:S55–61.
- [173] Dall'Igna OP, Porciuncula LO, Souza DO, Cunha RA, Lara DR. Neuroprotection by caffeine and adenosine A2A receptor blockade of  $\beta$ -amyloid neurotoxicity. *Br J Pharmacol* 2003;138:1207–9.
- [174] Fiebich BL, Biber K, Lieb K, van Calker D, Berger M, Bauer J, et al. Cyclooxygenase-2 expression in rat microglia is induced by adenosine A2a-receptors. *Glia* 1996;18:152–60.

- [175] Popoli P, Betto P, Reggio R, Ricciarello G. Adenosine A2A receptor stimulation enhances striatal extracellular glutamate levels in rats. *Eur J Pharmacol* 1995;287:215–7.
- [176] Popoli P, Pintor A, Domenici MR, Frank C, Tebano MT, Pezzola A, et al. Blockade of striatal adenosine A2A receptor reduces, through a presynaptic mechanism, quinolinic acid-induced excitotoxicity: possible relevance to neuroprotective interventions in neurodegenerative diseases of the striatum. *J Neurosci* 2002;22:1967–75.
- [177] Blum D, Galas MC, Pintor A, Brouillet E, Ledent C, Muller CE, et al. A dual role of adenosine A2A receptors in 3-nitropropionic acid-induced striatal lesions: implications for the neuroprotective potential of A2A antagonists. *J Neurosci* 2003;23:5361–9.
- [178] Tebano MT, Pintor A, Frank C, Domenici MR, Martire A, Pepponi R, et al. Adenosine A2A receptor blockade differentially influences excitotoxic mechanisms at pre- and postsynaptic sites in the rat striatum. *J Neurosci Res* 2004;77:100–7.
- [179] Xu K, Xu YH, Chen JF, Schwarzschild MA. Caffeine's neuroprotection against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity shows no tolerance to chronic caffeine administration in mice. *Neurosci Lett* 2002;322:13–6.
- [180] Waksman Y, Olson JM, Carlisle SJ, Cabral GA. The central cannabinoid receptor (CB1) mediates inhibition of nitric oxide production by rat microglial cells. *J Pharmacol Exp Ther* 1999;288:1357–66.
- [181] Molina-Holgado F, Pinteaux E, Moore JD, Molina-Holgado E, Guaza C, Gibson RM, et al. Endogenous interleukin-1 receptor antagonist mediates anti-inflammatory and neuroprotective actions of cannabinoids in neurons and glia. *J Neurosci* 2003;23:6470–4.
- [182] Ramirez BG, Blazquez C, Gomez DP, Guzman M, de Ceballos ML. Prevention of Alzheimer's disease pathology by cannabinoids: neuroprotection mediated by blockade of microglial activation. *J Neurosci* 2005;25:1904–13.
- [183] Abood ME, Rizvi G, Sallapudi N, McAllister SD. Activation of the CB1 cannabinoid receptor protects cultured mouse spinal neurons against excitotoxicity. *Neurosci Lett* 2001;309:197–201.
- [184] van der SM, Veldhuis WB, Bar PR, Veldink GA, Vliegthart JF, Nicolay K. Neuroprotection by Delta9-tetrahydrocannabinol, the main active compound in marijuana, against ouabain-induced in vivo excitotoxicity. *J Neurosci* 2001;21:6475–9.
- [185] Hansen HH, Azcoitia I, Pons S, Romero J, Garcia-Segura LM, Ramos JA, et al. Blockade of cannabinoid CB(1) receptor function protects against in vivo disseminating brain damage following NMDA-induced excitotoxicity. *J Neurochem* 2002;82:154–8.
- [186] van der SM, Veldhuis WB, Maccarrone M, Bar PR, Nicolay K, Veldink GA, et al. Acute neuronal injury, excitotoxicity, and the endocannabinoid system. *Mol Neurobiol* 2002;26:317–46.
- [187] van der SM, Hansen HH, Veldhuis WB, Bar PR, Nicolay K, Veldink GA, et al. Biosynthesis of endocannabinoids and their modes of action in neurodegenerative diseases. *Neurotox Res* 2003;5:183–200.
- [188] Marsicano G, Goodenough S, Monory K, Hermann H, Eder M, Cannich A, et al. CB1 cannabinoid receptors and on-demand defense against excitotoxicity. *Science* 2003;302:84–8.
- [189] Gerdeman G, Lovinger DM. CB1 cannabinoid receptor inhibits synaptic release of glutamate in rat dorsolateral striatum. *J Neurophysiol* 2001;85:468–71.
- [190] Azad SC, Eder M, Marsicano G, Lutz B, Zieglgansberger W, Rammes G. Activation of the cannabinoid receptor type 1 decreases glutamatergic and GABAergic synaptic transmission in the lateral amygdala of the mouse. *Learn Mem* 2003;10:116–28.
- [191] Melis M, Pistis M, Perra S, Muntoni AL, Pillolla G, Gessa GL. Endocannabinoids mediate presynaptic inhibition of glutamatergic transmission in rat ventral tegmental area dopamine neurons through activation of CB1 receptors. *J Neurosci* 2004;24:53–62.
- [192] Huang CC, Chen YL, Lo SW, Hsu KS. Activation of cAMP-dependent protein kinase suppresses the presynaptic cannabinoid inhibition of glutamatergic transmission at corticostriatal synapses. *Mol Pharmacol* 2002;61:578–85.
- [193] Dohovics R, Janaky R, Varga V, Hermann A, Saransaari P, Oja SS. Regulation of glutamatergic neurotransmission in the striatum by presynaptic adenylyl cyclase-dependent processes. *Neurochem Int* 2003;42:1–7.
- [194] Dohovics R, Janaky R, Varga V, Saransaari P, Oja SS. Cyclic AMP-mediated regulation of striatal glutamate release: interactions of presynaptic ligand- and voltage-gated ion channels and G-protein-coupled receptors. *Neurochem Int* 2003;43:425–30.
- [195] Rodriguez-Moreno A, Sihra TS. Presynaptic kainate receptor facilitation of glutamate release involves protein kinase A in the rat hippocampus. *J Physiol* 2004;557:733–45.
- [196] Iuvone T, Esposito G, Esposito R, Santamaria R, Di Rosa M, Izzo AA. Neuroprotective effect of cannabidiol, a non-psychoactive component from *Cannabis sativa*, on  $\beta$ -amyloid-induced toxicity in PC12 cells. *J Neurochem* 2004;89:134–41.
- [197] Raman C, McAllister SD, Rizvi G, Patel SG, Moore DH, Abood ME. Amyotrophic lateral sclerosis: delayed disease progression in mice by treatment with a cannabinoid. *Amyotroph Lateral Scler Other Motor Neuron Disord* 2004;5:33–9.
- [198] Nagayama T, Sinor AD, Simon RP, Chen J, Graham SH, Jin K, et al. Cannabinoids and neuroprotection in global and focal cerebral ischemia and in neuronal cultures. *J Neurosci* 1999;19:2987–95.
- [199] Lastres-Becker I, Bizat N, Boyer F, Hantraye P, Brouillet E, Fernandez-Ruiz J. Effects of cannabinoids in the rat model of Huntington's disease generated by an intrastriatal injection of malonate. *Neuroreport* 2003;14:813–6.
- [200] Semple DM, McIntosh AM, Lawrie SM. Cannabis as a risk factor for psychosis: systematic review. *J Psychopharmacol* 2005;19:187–94.
- [201] Bi XL, Yang JY, Dong YX, Wang JM, Cui YH, Ikeshima T, et al. Resveratrol inhibits nitric oxide and TNF- $\alpha$  production by lipopolysaccharide-activated microglia. *Int Immunopharmacol* 2005;5:185–93.
- [202] Wang MJ, Lin WW, Chen HL, Chang YH, Ou HC, Kuo JS, et al. Silymarin protects dopaminergic neurons against lipopolysaccharide-induced neurotoxicity by inhibiting microglia activation. *Eur J Neurosci* 2002;16:2103–12.
- [203] Li R, Huang YG, Fang D, Le WD. (-)-Epigallocatechin gallate inhibits lipopolysaccharide-induced microglial activation and protects against inflammation-mediated dopaminergic neuronal injury. *J Neurosci Res* 2004;78:723–31.
- [204] Lee H, Kim YO, Kim H, Kim SY, Noh HS, Kang SS, et al. Flavonoid wogonin from medicinal herb is neuroprotective by inhibiting inflammatory activation of microglia. *FASEB J* 2003;17:1943–4.



- [205] Hou RC, Chen HL, Tzen JT, Jeng KC. Effect of sesame antioxidants on LPS-induced NO production by BV2 microglial cells. *Neuroreport* 2003;14:1815–9.
- [206] Jeng KC, Hou RC, Wang JC, Ping LI. Sesamin inhibits lipopolysaccharide-induced cytokine production by suppression of p38 mitogen-activated protein kinase and nuclear factor- $\kappa$ B. *Immunol Lett* 2005;97:101–6.
- [207] Fujikawa T, Kanada N, Shimada A, Ogata M, Suzuki I, Hayashi I, et al. Effect of sesamin in *Acanthopanax senticosus* HARMS on behavioral dysfunction in rotenone-induced Parkinsonian rats. *Biol Pharm Bull* 2005;28:169–72.
- [208] Mattson MP, Lovell MA, Furukawa K, Markesbery WR. Neurotrophic factors attenuate glutamate-induced accumulation of peroxides, elevation of intracellular  $\text{Ca}^{2+}$  concentration, and neurotoxicity and increase antioxidant enzyme activities in hippocampal neurons. *J Neurochem* 1995;65:1740–51.
- [209] Duan W, Guo Z, Mattson MP. Brain-derived neurotrophic factor mediates an excitoprotective effect of dietary restriction in mice. *J Neurochem* 2001;76:619–26.
- [210] Cunningham LA, Su C. Astrocyte delivery of glial cell line-derived neurotrophic factor in a mouse model of Parkinson's disease. *Exp Neurol* 2002;174:230–42.
- [211] Mattson MP, Duan W, Guo Z. Meal size and frequency affect neuronal plasticity and vulnerability to disease: cellular and molecular mechanisms. *J Neurochem* 2003;84:417–31.
- [212] Duan W, Guo Z, Jiang H, Ware M, Mattson MP. Reversal of behavioral and metabolic abnormalities, and insulin resistance syndrome, by dietary restriction in mice deficient in brain-derived neurotrophic factor. *Endocrinology* 2003;144:2446–53.
- [213] Maswood N, Young J, Tilmont E, Zhang Z, Gash DM, Gerhardt GA, et al. Caloric restriction increases neurotrophic factor levels and attenuates neurochemical and behavioral deficits in a primate model of Parkinson's disease. *Proc Natl Acad Sci USA* 2004;101:18171–6.
- [214] Vaynman S, Ying Z, Gomez-Pinilla F. Hippocampal BDNF mediates the efficacy of exercise on synaptic plasticity and cognition. *Eur J Neurosci* 2004;20:2580–90.
- [215] Gobbo OL, O'Mara SM. Impact of enriched-environment housing on brain-derived neurotrophic factor and on cognitive performance after a transient global ischemia. *Behav Brain Res* 2004;152:231–41.
- [216] Laurin D, Verreault R, Lindsay J, MacPherson K, Rockwood K. Physical activity and risk of cognitive impairment and dementia in elderly persons. *Arch Neurol* 2001;58:498–504.
- [217] Luchsinger JA, Tang MX, Shea S, Mayeux R. Caloric intake and the risk of Alzheimer disease. *Arch Neurol* 2002;59:1258–63.
- [218] Wilson RS, Mendes De Leon CF, Barnes LL, Schneider JA, Bienias JL, Evans DA, et al. Participation in cognitively stimulating activities and risk of incident Alzheimer disease. *JAMA* 2002;287:742–8.
- [219] Wilson RS, Bennett DA, Bienias JL, Aggarwal NT, Mendes De Leon CF, Morris MC, et al. Cognitive activity and incident AD in a population-based sample of older persons. *Neurology* 2002;59:1910–4.
- [220] Chen H, Zhang SM, Schwarzschild MA, Hernan MA, Ascherio A. Physical activity and the risk of Parkinson disease. *Neurology* 2005;64:664–9.
- [221] Logroscino G, Marder K, Graziano J, Freyer G, Slavkovich V, Lojaciono N, et al. Dietary iron, animal fats, and risk of Parkinson's disease. *Mov Disord* 1998;13(Suppl. 1):13–6.
- [222] Johnson CC, Gorell JM, Rybicki BA, Sanders K, Peterson EL. Adult nutrient intake as a risk factor for Parkinson's disease. *Int J Epidemiol* 1999;28:1102–9.
- [223] McCarty MF. Does a vegan diet reduce risk for Parkinson's disease? *Med Hypotheses* 2001;57:318–23.
- [224] Ohta M, Mizuta I, Ohta K, Nishimura M, Mizuta E, Hayashi K, et al. Apomorphine up-regulates NGF and GDNF synthesis in cultured mouse astrocytes. *Biochem Biophys Res Commun* 2000;272:18–22.
- [225] Mizuta I, Ohta M, Ohta K, Nishimura M, Mizuta E, Kuno S. Riluzole stimulates nerve growth factor, brain-derived neurotrophic factor and glial cell line-derived neurotrophic factor synthesis in cultured mouse astrocytes. *Neurosci Lett* 2001;310:117–20.
- [226] Ohta K, Kuno S, Mizuta I, Fujinami A, Matsui H, Ohta M. Effects of dopamine agonists bromocriptine, pergolide, cabergoline, and SKF-38393 on GDNF, NGF, and BDNF synthesis in cultured mouse astrocytes. *Life Sci* 2003;73:617–26.
- [227] Toyomoto M, Inoue S, Ohta K, Kuno S, Ohta M, Hayashi K, et al. Production of NGF, BDNF and GDNF in mouse astrocyte cultures is strongly enhanced by a cerebral vasodilator, ifenprodil. *Neurosci Lett* 2005;379:185–9.
- [228] Koyama Y, Egawa H, Osakada M, Baba A, Matsuda T. Increase by FK960, a novel cognitive enhancer, in glial cell line-derived neurotrophic factor production in cultured rat astrocytes. *Biochem Pharmacol* 2004;68:275–82.
- [229] Palmer GC. Neuroprotection by NMDA receptor antagonists in a variety of neuropathologies. *Curr Drug Targets* 2001;2:241–71.
- [230] Sonkusare SK, Kaul CL, Ramarao P. Dementia of Alzheimer's disease and other neurodegenerative disorders – memantine, a new hope. *Pharmacol Res* 2005;51:1–17.
- [231] Lipton SA. Paradigm shift in NMDA receptor antagonist drug development: molecular mechanism of uncompetitive inhibition by memantine in the treatment of Alzheimer's disease and other neurologic disorders. *J Alzheimers Dis* 2004;6:S61–74.
- [232] Molinuevo JL, Llado A, Rami L. Memantine: targeting glutamate excitotoxicity in Alzheimer's disease and other dementias. *Am J Alzheimers Dis Other Demen* 2005;20:77–85.
- [233] Gredal O, Werdelin L, Bak S, Christensen PB, Boysen G, Kristensen MO, et al. A clinical trial of dextromethorphan in amyotrophic lateral sclerosis. *Acta Neurol Scand* 1997;96:8–13.
- [234] Ooboshi H, Sadoshima S, Yao H, Ibayashi S, Matsumoto T, Uchimura H, et al. Ischemia-induced release of amino acids in the hippocampus of aged hypertensive rats. *J Cereb Blood Flow Metab* 1995;15:227–34.
- [235] Phillis JW, Song D, O'Regan MH. Inhibition by anion channel blockers of ischemia-evoked release of excitotoxic and other amino acids from rat cerebral cortex. *Brain Res* 1997;758:9–16.
- [236] Saransaari P, Oja SS. Mechanisms of ischemia-induced taurine release in mouse hippocampal slices. *Brain Res* 1998;807:118–24.
- [237] Saransaari P, Oja SS. Taurine and neural cell damage. *Amino Acids* 2000;19:509–26.
- [238] Bureau MH, Olsen RW. Taurine acts on a subclass of GABAA receptors in mammalian brain in vitro. *Eur J Pharmacol* 1991;207:9–16.
- [239] Dawson Jr R, Liu S, Eppler B, Patterson T. Effects of dietary taurine supplementation or deprivation in aged male Fischer 344 rats. *Mech Ageing Dev* 1999;107:73–91.
- [240] Zhao P, Huang YL, Cheng JS. Taurine antagonizes calcium overload induced by glutamate or chemical hypoxia in

- cultured rat hippocampal neurons. *Neurosci Lett* 1999;268:25–8.
- [241] O'Byrne MB, Tipton KF. Taurine-induced attenuation of MPP<sup>+</sup> neurotoxicity in vitro: a possible role for the GABA(A) subclass of GABA receptors. *J Neurochem* 2000;74:2087–93.
- [242] Louzada PR, Lima AC, Mendonca-Silva DL, Noel F, De Mello FG, Ferreira ST. Taurine prevents the neurotoxicity of  $\beta$ -amyloid and glutamate receptor agonists: activation of GABA receptors and possible implications for Alzheimer's disease and other neurological disorders. *FASEB J* 2004;18:511–8.
- [243] Rivas-Arancibia S, Rodriguez AI, Zigova T, Willing AE, Brown WD, Cahill DW, et al. Taurine increases rat survival and reduces striatal damage caused by 3-nitropropionic acid. *Int J Neurosci* 2001;108:55–67.
- [244] Guo J, Li R, Zhao P, Cheng J. Effect of taurine in combination with electroacupuncture on neuronal damage following transient focal cerebral ischemia in rats. *Acupunct Electrother Res* 2002;27:129–36.
- [245] Shuaib A. The role of taurine in cerebral ischemia: studies in transient forebrain ischemia and embolic focal ischemia in rodents. *Adv Exp Med Biol* 2003;526:421–31.
- [246] Durelli L, Mutani R. The current status of taurine in epilepsy. *Clin Neuropharmacol* 1983;6:37–48.
- [247] Fariello RG, Golden GT, McNeal Jr RB. Taurine and related amino acids in seizure disorders – current controversies. *Prog Clin Biol Res* 1985;179:413–24.
- [248] Kearns S, Dawson Jr R. Cytoprotective effect of taurine against hypochlorous acid toxicity to PC12 cells. *Adv Exp Med Biol* 2000;483:563–70.
- [249] Serban V, Liu Y, Quinn MR. Production of nitric oxide by activated microglial cells is inhibited by taurine chloramine. *Adv Exp Med Biol* 2003;526:357–64.
- [250] Jussofie A, Schmitz A, Hiemke C. Kavapyrone enriched extract from Piper methysticum as modulator of the GABA binding site in different regions of rat brain. *Psychopharmacology (Berl)* 1994;116:469–74.
- [251] Backhaus C, Krieglstein J. Extract of kava (Piper methysticum) and its methysticin constituents protect brain tissue against ischemic damage in rodents. *Eur J Pharmacol* 1992;215:265–9.
- [252] Mattson MP, Duan W, Wan R, Guo Z. Prophylactic activation of neuroprotective stress response pathways by dietary and behavioral manipulations. *NeuroRx* 2004;1:111–6.
- [253] Lee J, Herman JP, Mattson MP. Dietary restriction selectively decreases glucocorticoid receptor expression in the hippocampus and cerebral cortex of rats. *Exp Neurol* 2000;166:435–41.
- [254] Flier J, Van Muiswinkel FL, Jongenelen CA, Drukarch B. The neuroprotective antioxidant  $\alpha$ -lipoic acid induces detoxication enzymes in cultured astroglial cells. *Free Radic Res* 2002;36:695–9.
- [255] McCarty MF. Versatile cytoprotective activity of lipoic acid may reflect its ability to activate signalling intermediates that trigger the heat-shock and phase II responses. *Med Hypotheses* 2001;57:313–7.
- [256] Suh JH, Shenvi SV, Dixon BM, Liu H, Jaiswal AK, Liu RM, et al. Decline in transcriptional activity of Nrf2 causes age-related loss of glutathione synthesis, which is reversible with lipoic acid. *Proc Natl Acad Sci USA* 2004;101:3381–6.
- [257] Wolz P, Krieglstein J. Neuroprotective effects of  $\alpha$ -lipoic acid and its enantiomers demonstrated in rodent models of focal cerebral ischemia. *Neuropharmacology* 1996;35:369–75.
- [258] Panigrahi M, Sadguna Y, Shivakumar BR, Kolluri SV, Roy S, Packer L, et al.  $\alpha$ -Lipoic acid protects against reperfusion injury following cerebral ischemia in rats. *Brain Res* 1996;717:184–8.
- [259] Packer L, Tritschler HJ, Wessel K. Neuroprotection by the metabolic antioxidant  $\alpha$ -lipoic acid. *Free Radic Biol Med* 1997;22:359–78.
- [260] Andreassen OA, Dedeoglu A, Friedlich A, Ferrante KL, Hughes D, Szabo C, et al. Effects of an inhibitor of poly(ADP-ribose) polymerase, desmethylselegiline, trientine, and lipoic acid in transgenic ALS mice. *Exp Neurol* 2001;168:419–24.
- [261] Zhang L, Xing GQ, Barker JL, Chang Y, Maric D, Ma W, et al.  $\alpha$ -Lipoic acid protects rat cortical neurons against cell death induced by amyloid and hydrogen peroxide through the Akt signalling pathway. *Neurosci Lett* 2001;312:125–8.
- [262] Andreassen OA, Ferrante RJ, Dedeoglu A, Beal MF. Lipoic acid improves survival in transgenic mouse models of Huntington's disease. *Neuroreport* 2001;12:3371–3.
- [263] Bharat S, Cochran BC, Hsu M, Liu J, Ames BN, Andersen JK. Pre-treatment with R-lipoic acid alleviates the effects of GSH depletion in PC12 cells: implications for Parkinson's disease therapy. *Neurotoxicology* 2002;23:479–86.
- [264] Ziegler D, Reljanovic M, Mehnert H, Gries FA.  $\alpha$ -Lipoic acid in the treatment of diabetic polyneuropathy in Germany: current evidence from clinical trials. *Exp Clin Endocrinol Diabetes* 1999;107:421–30.
- [265] Savaskan NE, Brauer AU, Kuhbacher M, Eyupoglu IY, Kyriakopoulos A, Ninnemann O, et al. Selenium deficiency increases susceptibility to glutamate-induced excitotoxicity. *FASEB J* 2003;17:112–4.
- [266] Chen J, Berry MJ. Selenium and selenoproteins in the brain and brain diseases. *J Neurochem* 2003;86:1–12.
- [267] Schweizer U, Brauer AU, Kohrte J, Nitsch R, Savaskan NE. Selenium and brain function: a poorly recognized liaison. *Brain Res Brain Res Rev* 2004;45:164–78.
- [268] Brauer AU, Savaskan NE. Molecular actions of selenium in the brain: neuroprotective mechanisms of an essential trace element. *Rev Neurosci* 2004;15:19–32.
- [269] Christen S, Woodall AA, Shigenaga MK, Southwell-Keely PT, Duncan MW, Ames BN.  $\gamma$ -tocopherol traps mutagenic electrophiles such as NO(X) and complements  $\alpha$ -tocopherol: physiological implications. *Proc Natl Acad Sci USA* 1997;94:3217–22.
- [270] Jiang Q, Lykkesfeldt J, Shigenaga MK, Shigeno ET, Christen S, Ames BN.  $\gamma$ -Tocopherol supplementation inhibits protein nitration and ascorbate oxidation in rats with inflammation. *Free Radic Biol Med* 2002;33:1534–42.
- [271] Matthews RT, Yang L, Jenkins BG, Ferrante RJ, Rosen BR, Kaddurah-Daouk R, et al. Neuroprotective effects of creatine and cyclocreatine in animal models of Huntington's disease. *J Neurosci* 1998;18:156–63.
- [272] Malcon C, Kaddurah-Daouk R, Beal MF. Neuroprotective effects of creatine administration against NMDA and malonate toxicity. *Brain Res* 2000;860:195–8.
- [273] Beal MF. Bioenergetic approaches for neuroprotection in Parkinson's disease. *Ann Neurol* 2003;53(Suppl. 3):S39–47.
- [274] Zhu S, Li M, Figueroa BE, Liu A, Stavrovskaya IG, Pasinelli P, et al. Prophylactic creatine administration mediates neuroprotection in cerebral ischemia in mice. *J Neurosci* 2004;24:5909–12.
- [275] Schulz JB, Henshaw DR, Matthews RT, Beal MF. Coenzyme Q10 and nicotinamide and a free radical spin trap protect against MPTP neurotoxicity. *Exp Neurol* 1995;132:279–83.

- [276] Shults CW, Oakes D, Kiebertz K, Beal MF, Haas R, Plumb S, et al. Effects of coenzyme Q10 in early Parkinson disease: evidence of slowing of the functional decline. *Arch Neurol* 2002;59:1541–50.
- [277] Beal MF. Mitochondrial dysfunction and oxidative damage in Alzheimer's and Parkinson's diseases and coenzyme Q10 as a potential treatment. *J Bioenerg Biomembr* 2004;36:381–6.
- [278] Virmani MA, Biselli R, Spadoni A, Rossi S, Corsico N, Calvani M, et al. Protective actions of L-carnitine and acetyl-L-carnitine on the neurotoxicity evoked by mitochondrial uncoupling or inhibitors. *Pharmacol Res* 1995;32:383–9.
- [279] Veech RL, Chance B, Kashiwaya Y, Lardy HA, Cahill Jr GF. Ketone bodies, potential therapeutic uses. *IUBMB Life* 2001;51:241–7.
- [280] Tieu K, Perier C, Caspersen C, Teismann P, Wu DC, Yan SD, et al. D-β-hydroxybutyrate rescues mitochondrial respiration and mitigates features of Parkinson disease. *J Clin Invest* 2003;112:892–901.
- [281] Massieu L, Haces ML, Montiel T, Hernandez-Fonseca K. Acetoacetate protects hippocampal neurons against glutamate-mediated neuronal damage during glycolysis inhibition. *Neuroscience* 2003;120:365–78.
- [282] Reger MA, Henderson ST, Hale C, Cholerton B, Baker LD, Watson GS, et al. Effects of β-hydroxybutyrate on cognition in memory-impaired adults. *Neurobiol Aging* 2004;25:311–4.
- [283] de la Torre JC. Cerebral hypoperfusion, capillary degeneration, and development of Alzheimer disease. *Alzheimer Dis Assoc Disord* 2000;14(Suppl. 1):S72–81.
- [284] Lindeberg S. Apparent absence of cerebrovascular disease in Melanesians. PhD Thesis, Lund University, Sweden; 1994.
- [285] Lindeberg S, Lundh B. Apparent absence of stroke and ischaemic heart disease in a traditional Melanesian island: a clinical study in Kitava. *J Intern Med* 1993;233:269–75.
- [286] McCarty MF. Up-regulation of endothelial nitric oxide activity as a central strategy for prevention of ischemic stroke – just say NO to stroke!. *Med Hypotheses* 2000;55:386–403 [JID – 7505668].
- [287] Trowell HC. Hypertension, obesity, diabetes mellitus and coronary heart disease. In: Trowell HC, Burkitt DP, editors. *Western diseases: their emergence and prevention*. Cambridge, MA: Harvard University Press; 1981. p. 3–32.
- [288] Gustafson D, Rothenberg E, Blennow K, Steen B, Skoog I. An 18-year follow-up of overweight and risk of Alzheimer disease. *Arch Intern Med* 2003;163:1524–8.
- [289] Luchsinger JA, Tang MX, Shea S, Mayeux R. Hyperinsulinemia and risk of Alzheimer disease. *Neurology* 2004;63:1187–92.
- [290] Watson GS, Peskind ER, Asthana S, Purganan K, Wait C, Chapman D, et al. Insulin increases CSF Abeta42 levels in normal older adults. *Neurology* 2003;60:1899–903.
- [291] Morikawa E, Huang Z, Moskowitz MA. L-arginine decreases infarct size caused by middle cerebral arterial occlusion in SHR. *Am J Physiol* 1992;263:H1632–5.
- [292] Milstien S, Katusic Z. Oxidation of tetrahydrobiopterin by peroxynitrite: implications for vascular endothelial function. *Biochem Biophys Res Commun* 1999;263:681–4.
- [293] Stroes ES, van Faassen EE, Yo M, Martasek P, Boer P, Govers R, et al. Folic acid reverts dysfunction of endothelial nitric oxide synthase. *Circ Res* 2000;86:1129–34.
- [294] Hyndman ME, Verma S, Rosenfeld RJ, Anderson TJ, Parsons HG. Interaction of 5-methyltetrahydrofolate and tetrahydrobiopterin on endothelial function. *Am J Physiol Heart Circ Physiol* 2002;282:H2167–72.
- [295] Verhaar MC, Stroes E, Rabelink TJ. Folates and cardiovascular disease. *Arterioscler Thromb Vasc Biol* 2002;22:6–13.
- [296] McCarty MF. Coping with endothelial superoxide: potential complementarity of arginine and high-dose folate. *Med Hypotheses* 2004;63:709–18.
- [297] Quadri P, Fragiaco C, Pezzati R, Zanda E, Forloni G, Tettamanti M, et al. Homocysteine, folate, and vitamin B-12 in mild cognitive impairment, Alzheimer disease, and vascular dementia. *Am J Clin Nutr* 2004;80:114–22.
- [298] Wald DS, Law M, Morris JK. Homocysteine and cardiovascular disease: evidence on causality from a meta-analysis. *BMJ* 2002;325:1202.
- [299] Klivenyi P, Gardian G, Calingasan NY, Yang L, Beal MF. Additive neuroprotective effects of creatine and a cyclooxygenase 2 inhibitor against dopamine depletion in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) mouse model of Parkinson's disease. *J Mol Neurosci* 2003;21:191–8.
- [300] Klivenyi P, Kiaei M, Gardian G, Calingasan NY, Beal MF. Additive neuroprotective effects of creatine and cyclooxygenase 2 inhibitors in a transgenic mouse model of amyotrophic lateral sclerosis. *J Neurochem* 2004;88:576–82.

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